

## TLA-110 Rotor



**Used In Beckman Coulter  
Optima™ MAX-XP, MAX, MAX-E,  
TL, TLX, and TL-100  
Tabletop Ultracentrifuges**



## SAFETY NOTICE

This safety notice summarizes information basic to the safe use of the rotor described in this manual. The international symbol displayed above is a reminder to the user that all safety instructions should be read and understood before operation or maintenance of this equipment is attempted. When you see the symbol on other pages throughout this publication, pay special attention to the specific safety information presented. Observance of safety precautions will also help to avoid actions that could damage or adversely affect the performance of the rotor. This rotor was developed, manufactured, and tested for safety and reliability as part of a Beckman Coulter ultracentrifuge/rotor system. Its safety or reliability cannot be assured if used in a centrifuge not of Beckman Coulter's manufacture or in a Beckman Coulter ultracentrifuge that has been modified without Beckman Coulter's approval.



Handle body fluids with care because they can transmit disease. No known test offers complete assurance that such fluids are free of micro-organisms. Some of the most virulent—Hepatitis (B and C) viruses, HIV (I–V), atypical mycobacteria, and certain systemic fungi—further emphasize the need for aerosol protection. Handle other infectious samples according to good laboratory procedures and methods to prevent spread of disease. Because spills may generate aerosols, observe proper safety precautions for aerosol containment. Do not run toxic, pathogenic, or radioactive materials in this rotor without taking appropriate safety precautions. Biosafe containment should be used when Risk Group II materials (as identified in the World Health Organization *Laboratory Biosafety Manual*) are handled; materials of a higher group require more than one level of protection.



The rotor and accessories are not designed for use with materials capable of developing flammable or explosive vapors. Do not centrifuge such materials in nor handle or store them near the ultracentrifuge.



Although rotor components and accessories made by other manufacturers may fit in the TLA-110 rotor, their safety in this rotor cannot be ascertained by Beckman Coulter. Use of other manufacturers' components or accessories in the TLA-110 rotor may void the rotor warranty and should be prohibited by your laboratory safety officer. Only the components and accessories listed in this publication should be used in this rotor.



Make sure that filled containers are loaded symmetrically into the rotor and that opposing tubes are filled to the same level with liquid of the same density. Make sure that cavities in use have the proper spacers and/or floating spacers inserted before installing the rotor lid.



If disassembly reveals evidence of leakage, you should assume that some fluid escaped the rotor. Apply appropriate decontamination procedures to the centrifuge and accessories.

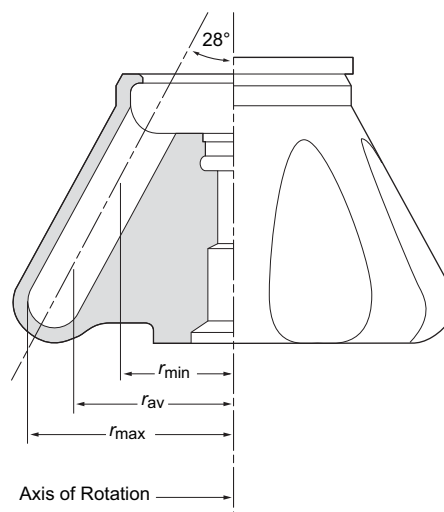


Never exceed the maximum rated speed of the rotor and labware in use. Refer to the section on RUN SPEEDS, and derate the run speed as appropriate.



Do not use sharp tools on the rotor that could cause scratches in the rotor surface. Corrosion begins in scratches and may open fissures in the rotor with continued use.

## TLA-110 ROTOR



## SPECIFICATIONS

Maximum speed	110 000 rpm
Density rating at maximum speed	1.7 g/mL
Relative Centrifugal Field* at maximum speed	
At $r_{\max}$ (48.5 mm)	$657\,000 \times g$
At $r_{\text{av}}$ (37.2 mm)	$504\,000 \times g$
At $r_{\min}$ (26.0 mm)	$353\,000 \times g$
$k$ factor at maximum speed	13
Conditions requiring speed reductions	see RUN SPEEDS
Number of tube cavities	8
Available tubes	see Table 1
Nominal tube dimensions (largest tube)	13 × 51 mm
Nominal tube capacity	5.1 mL
Nominal rotor capacity	40.8 mL
Approximate acceleration time to maximum	
speed (fully loaded)	4 min
Approximate deceleration time from maximum	
speed (fully loaded)	2 min
Weight of fully loaded rotor	1.0 kg (2.2 lb)
Rotor material	titanium

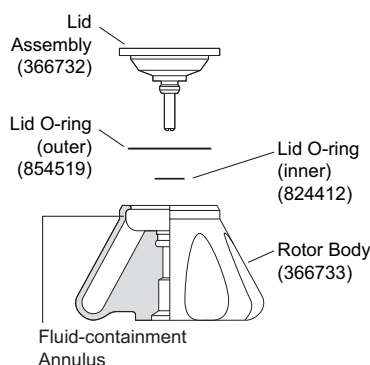
\* Relative Centrifugal Field (RCF) is the ratio of the centrifugal acceleration at a specified radius and speed ( $r\omega^2$ ) to the standard acceleration of gravity ( $g$ ) according to the following formula:

$$\text{RCF} = \frac{r\omega^2}{g}$$

where  $r$  is the radius in millimeters,  $\omega$  is the angular velocity in radians per second ( $2\pi \text{ RPM} / 60$ ), and  $g$  is the standard acceleration of gravity (9807 mm/s<sup>2</sup>). After substitution:

$$\text{RCF} = 1.12 r \left( \frac{\text{RPM}}{1000} \right)^2$$

## DESCRIPTION



*This Beckman Coulter rotor has been manufactured in a registered ISO 9001 or 9002 facility for use with the appropriately classified Beckman Coulter ultracentrifuge.*

The TLA-110 fixed angle rotor, rated for 110 000 rpm, has a tube angle of 28 degrees from the axis of rotation. The rotor can centrifuge up to eight tubes and is used in Beckman Coulter Optima™ MAX, MAX-XP, MAX-E, TL, TLX, and the TL-100 tabletop ultracentrifuges.

The rotor is made of titanium and is finished with black polyurethane paint. The lid is made of aluminum and anodized to resist corrosion. A plunger in the lid locks the rotor to the drive hub before beginning the run, and two lubricated O-rings made of Buna-N rubber maintain atmospheric pressure inside the rotor during centrifugation. The tube cavities are numbered to aid in sample identification.

The rotor is specially designed with a fluid-containment annulus located below the O-ring sealing surface. The annulus retains fluid that may escape from leaking or overfilled tubes, thereby preventing the liquid from escaping into the instrument chamber.

The ultracentrifuge identifies rotor speed during the run by means of a magnetic speed sensor in the instrument chamber and magnets on the bottom of the rotor. This overspeed protection system ensures that the rotor does not exceed its maximum permitted speed.

This rotor was tested\* to demonstrate containment of microbiological aerosols under normal operating conditions of the associated Beckman Coulter centrifuge, when used and maintained as instructed.

See the Warranty at the back of this manual for warranty information.

## PREPARATION AND USE

*Specific information about the TLA-110 rotor is given here. Information common to this and other rotors is contained in Rotors and Tubes for Tabletop Preparative Ultracentrifuges (publication TLR-IM), which should be used together with this manual for complete rotor and accessory operation. Publication TLR-IM is included in the literature package shipped with the rotor.*

\* Validation of microbiological containment was done at an independent third-party testing facility (CAMR, Porton Down, UK, or USAMRIID, Ft. Detrick, MD, U.S.A.). Improper use or maintenance may affect seal integrity and thus containment.

**NOTE**

Although rotor components and accessories made by other manufacturers may fit in the TLA-110 rotor, their safety in this rotor cannot be ascertained by Beckman Coulter. Use of other manufacturers' components or accessories in the TLA-110 rotor may void the rotor warranty and should be prohibited by your laboratory safety officer. Only the components and accessories listed in this publication should be used in this rotor.

**PRERUN SAFETY CHECKS**

*Read the Safety Notice page at the front of this manual before using the rotor.*

1. Inspect the O-rings and plunger mechanism for damage—the high forces generated in this rotor can cause damaged components to fail.
2. Use only tubes and accessories listed in Table 1.
3. Check the chemical compatibilities of all materials used (refer to Appendix A in *Rotors and Tubes*).

**ROTOR PREPARATION**

*For runs at other than room temperature, refrigerate or warm the rotor beforehand for fast equilibration.*

1. Lightly but evenly lubricate metal threads with Spinkote™ lubricant (306812).
2. Apply a thin film of silicone vacuum grease (335148) to the two O-rings in the rotor lid.
3. If using 1.5 mL Microfuge® tubes, place the adapters into the tube cavities before inserting the tubes. Cut the hinges off the caps. Face the lift tab of all tubes toward the outside of the rotor (see Figure 1) before installing the rotor lid.

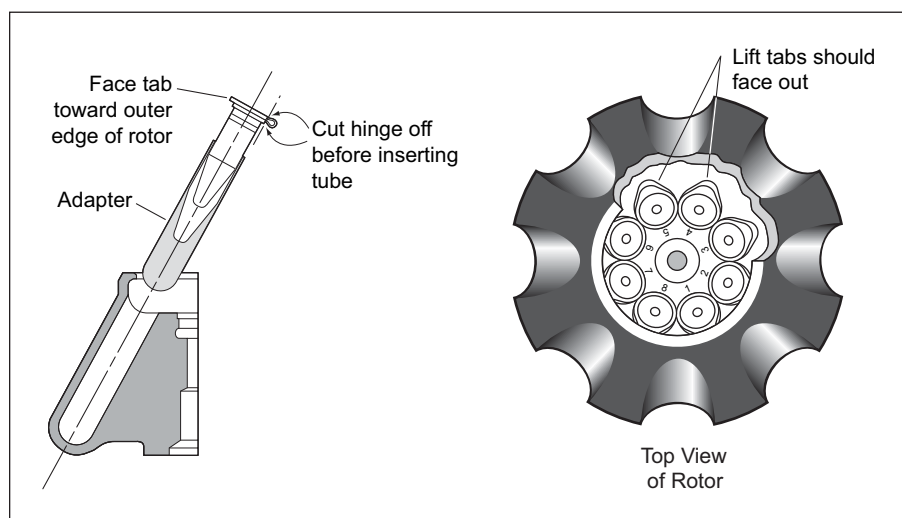
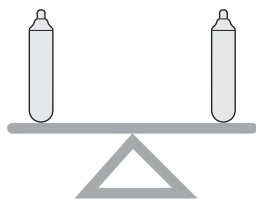


Figure 1. Tube Cap Orientation for Microfuge Tubes



4. Load the filled and capped tubes symmetrically into the rotor (see page 8 for tube information). If fewer than eight tubes are being run, they must be arranged symmetrically in the rotor (see Figure 2). *Opposing tubes must be filled to the same level with liquid of the same density.*
5. Use the required spacers, if necessary (see Table 1), to complete the loading operation.
6. After the rotor is loaded, insert it into the portable polypropylene rotor vise (346133). Place the lid on the rotor and tighten it firmly to the right (clockwise) by hand. No tool is required.

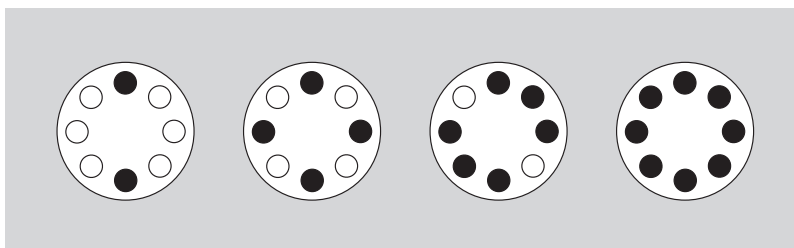
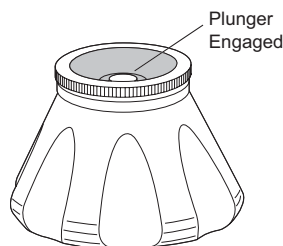


Figure 2. Arranging Tubes in the Rotor. Two, four, six, or eight tubes can be centrifuged per run if they are arranged in the rotor as shown.

## OPERATION

---



1. Use an absorbent towel to wipe off condensation from the rotor, then carefully place the rotor on the drive hub.
2. Lock the rotor in place by gently pressing the plunger down until you feel it click. When you remove your finger, the plunger will remain flush with the rotor body if it is properly engaged. If the plunger pops up, repeat the procedure. (The Optima MAX, MAX-XP, and MAX-E ultracentrifuges automatically secure the rotor to the drive shaft without the need for engaging the plunger.)



### CAUTION

---

*In all ultracentrifuge models except the Optima MAX, MAX-XP, and MAX-E, it is very important to lock the rotor in place before beginning the run to ensure that the rotor remains seated during centrifugation. Failure to lock the rotor in place before beginning the run may result in damage to both rotor and instrument.*

---

3. Refer to the instrument instruction manual for ultracentrifuge operation.
4. For additional operating information, see the following:
  - RUN TIMES, page 11, for using  $k$  factors to adjust run durations.
  - RUN SPEEDS, page 12, for information about speed limitations.
  - SELECTING CsCl GRADIENTS, page 14, for methods to avoid CsCl precipitation during centrifugation.

## REMOVAL AND SAMPLE RECOVERY

---

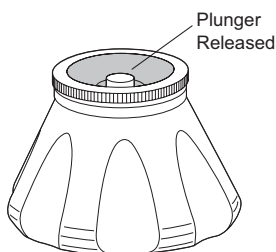


### CAUTION

---

*If disassembly reveals evidence of leakage, you should assume that some fluid escaped the rotor. Apply appropriate decontamination procedures to the centrifuge and accessories.*

---



1. To release the plunger at the end of the run, gently press it down until you feel it click. When you remove your finger the plunger will pop up to its released position.
2. Remove the rotor from the ultracentrifuge and place it in the rotor vise.
3. Remove the lid by unscrewing it to the left (counterclockwise).
4. Use a tube removal tool to remove the spacers and tubes.

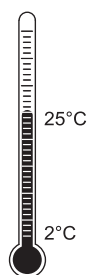
## TUBES AND ACCESSORIES

---

The TLA-110 rotor uses tubes and accessories listed in Table 1. Be sure to use only those items listed, and to observe the maximum speed limits shown. Refer to Appendix A in *Rotors and Tubes* for information on the chemical resistances of tube and accessory materials.

### Temperature Limits

---

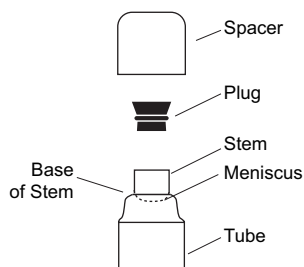


- Plastic tubes have been centrifuge tested for use at temperatures between 2 and 25°C. For centrifugation at other temperatures, pretest tubes under anticipated run conditions.
- If plastic containers are frozen before use, make sure that they are thawed to at least 2°C prior to centrifugation.

### OptiSeal™ Tubes

---

OptiSeal tubes come with plastic plugs and can be quickly and easily prepared for use. With the tube spacer in place, the *g* force during centrifugation ensures a tight, reliable seal that protects your samples.



- Place the tubes in the rack and fill each tube to the base of the stem, leaving no fluid in the stem. Overfilling the tube can cause spillage when the plug is inserted or can compromise seal integrity. However, too much air can cause excessive tube deformation, disrupting gradients and sample bands.



*Table 1. Available Tubes for the TLA-110 Rotor.  
Use only the items listed here and observe maximum fill volumes and speed shown.*

Tube			Required Accessory		Max. Speed/ RCF/ k Factor
Dimensions and Volume	Description	Part Number	Description	Part Number	
13 × 51 mm 5.1 mL	Quick-Seal polyallomer, bell top	362248 (pkg/50)	Delrin <sup>a</sup> floating spacer	362307 (pkg/8)	110 000 rpm 657 000 × <i>g</i> 13
13 × 48 mm 4.7 mL	OptiSeal <sup>b</sup> bell-top	361621 (pkg/56)	Ultem <sup>c</sup> spacer	361676 (pkg/2)	110 000 rpm 657 000 × <i>g</i> 12
13 × 32 mm 3.5 mL	Quick-Seal polyallomer, bell-top	349621 (pkg/50)	Noryl <sup>c</sup> floating spacer	360270 (pkg/8)	110 000 rpm 657 000 × <i>g</i> 7
13 × 56 mm 3.2 mL <sup>d</sup>	thickwall polycarbonate	362305 (pkg/50)	none	—	110 000 rpm 657 000 × <i>g</i> 15
13 × 56 mm 3.2 mL	thickwall polyallomer	362333 (pkg/50)	none	—	70 000 rpm 266 000 × <i>g</i> 37
13 × 25 mm 2.0 mL	Quick-Seal polyallomer, bell-top	345829 (pkg/50)	Noryl floating spacer	360270 (pkg/8)	110 000 rpm 657 000 × <i>g</i> 5
11 × 39 mm 1.5 mL	Microfuge, polyallomer (capped)	357448 (pkg/500)	Delrin adapter	360951 (pkg/8)	70 000 rpm <sup>e</sup> 206 000 × <i>g</i> 19
11 × 47 mm 1.5 mL	Microfuge, polypropylene (capped)	Corning 430909 <sup>f</sup> (pkg/500)	Delrin adapter	393238 (pkg/8)	110 000 rpm 657 000 × <i>g</i> 9

<sup>a</sup> Delrin is a registered trademark of E.I. Du Pont de Nemours & Company.

<sup>b</sup> Includes disposable plastic plugs.

<sup>c</sup> Ultem and Noryl are registered trademarks of GE Plastics.

<sup>d</sup> Minimum fill level for this tube is 1.6 mL.

<sup>e</sup> At 40°C, speed must be reduced to 59 000 rpm.

<sup>f</sup> The orange plastic piece that attaches the cap to the tube must be removed or the tube will not seat properly in the adapter or in the rotor. Use scissors to cut the plastic piece at the cap and at the tube body. The orange ring around the tube can be left in place.

- Refer to *Using OptiSeal™ Tubes* (publication IN-189), included in each box of tubes, for detailed information on the use and care of OptiSeal tubes.



## Quick-Seal® Tubes

---

Quick-Seal tubes must be sealed prior to centrifugation. These tubes are heat sealed and do not need caps; however, spacers are required on top of the tubes when they are loaded into the rotor.

- Fill Quick-Seal tubes leaving a *small* bubble of air at the base of the neck. Do not leave a large air space—too much air can cause excessive tube deformation.
- Refer to *Rotors and Tubes* for detailed information on the use and care of Quick-Seal tubes.

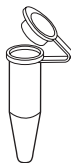
Some of the tubes listed in Table 1 are part of the g-Max™ system. The g-Max system uses a combination of small bell-top Quick-Seal tubes and floating spacers (also called g-Max spacers). This means that you can run the shorter tubes listed in the table in the TLA-110 rotor without reduction in *g* force. Additional information about the g-Max system is available in publication DS-709.

## Thickwall Tubes

---



Thickwall polyallomer and polycarbonate tubes can be run partially filled (at least half filled) without caps, but all opposing tubes for a run must be filled to the same level with liquid of the same density. Do not overfill capless tubes; be sure to note the reduction in run speed shown in Table 1.

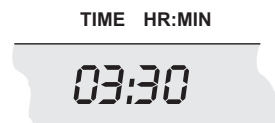


## Microfuge Tubes

---

The 1.5-mL microfuge tubes, with attached caps, are made of clear polyallomer. The tubes are placed in adapters for use in this rotor. All opposing tubes for a run must be filled with liquid of the same density. Be sure to note the run speed reduction shown in Table 1.

## RUN TIMES



The  $k$  factor of the rotor is a measure of the rotor's pelleting efficiency. (Beckman Coulter has calculated the  $k$  factors for all of its preparative rotors at maximum rated speed and using full tubes.) The  $k$  factor is calculated from the formula:

$$k = \frac{\ln(r_{\max}/r_{\min})}{\omega^2} \times \frac{10^{13}}{3600} \quad (1)$$

where  $\omega$  is the angular velocity of the rotor in radians per second ( $\omega = 0.105 \times \text{rpm}$ ),  $r_{\max}$  is the maximum radius, and  $r_{\min}$  is the minimum radius.

After substitution:

$$k = \frac{(2.533 \times 10^{11}) \ln(r_{\max}/r_{\min})}{\text{rpm}^2} \quad (2)$$

Use the  $k$  factor in the following equation to estimate the run time  $t$  (in hours) required to pellet particles of known sedimentation coefficient  $s$  (in Svedberg units,  $S$ ).

$$t = \frac{k}{s} \quad (3)$$

Run times can be estimated for centrifugation at less than maximum speed by adjusting the  $k$  factor as follows:

$$k_{\text{adj}} = k \left( \frac{110\,000}{\text{actual run speed}} \right)^2 \quad (4)$$

Run times can also be estimated from data established in prior experiments if the  $k$  factor of the previous rotor is known. For any two rotors, a and b:

$$\frac{t_a}{t_b} = \frac{k_a}{k_b} \quad (5)$$

For more information on  $k$  factors see *Use of  $k$  Factor for Estimating Run Times from Previously Established Run Conditions* (publication DS-719).

## RUN SPEEDS

---



The centrifugal force at a given radius in a rotor is a function of speed. Comparisons of forces between different rotors are made by comparing the rotors' relative centrifugal fields (RCF). When rotational speed is adjusted so that identical samples are subjected to the same RCF in two different rotors, the samples are subjected to the same force. The RCF at a number of rotor speeds is provided in Tables 2 and 3.

Speeds must be reduced under the following circumstances:

1. If nonprecipitating solutions more dense than 1.7 g/mL are centrifuged, the maximum allowable run speed must be reduced according to the following equation:

$$\text{reduced maximum speed} = (110\,000 \text{ rpm}) \sqrt{\frac{1.7 \text{ g/mL}}{\rho}} \quad (6)$$

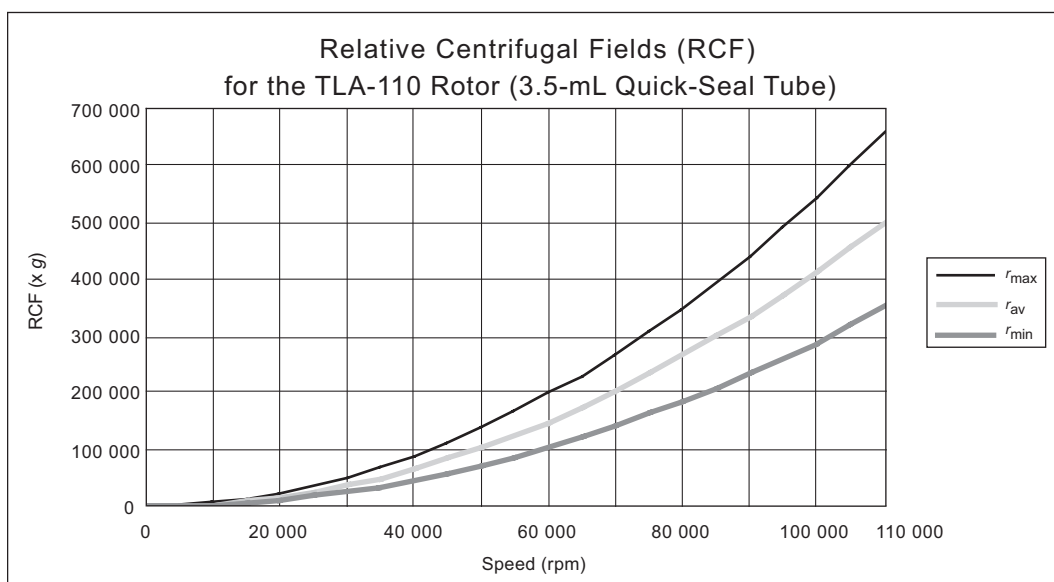
where  $\rho$  is the density of the tube contents. This speed reduction will protect the rotor from excessive stresses due to the added tube load. *Note, however, that the use of this formula may still produce maximum speed values that are higher than the limitations imposed by the use of certain tubes or adapters.* In such cases, use the lower of the two values.

2. *Further speed limits must be imposed* when CsCl or other self-forming-gradient salts are centrifuged, as equation (6) does not predict concentration limits/speeds that are required to avoid precipitation of salt crystals. Precipitation during centrifugation would alter the density distribution of CsCl and this would change the position of the sample bands. Figures 3 and 4, together with the description and examples below, show how to reduce run speeds when using CsCl gradients.

*Table 2. Relative Centrifugal Fields for the TLA-110 Rotor  
when Using g-Max™ 3.5-mL Quick-Seal® Tubes.  
Entries in this table are calculated from the formula  
 $RCF = 1.12r (RPM/1000)^2$  and then rounded to three significant digits.*

Rotor Speed (rpm)	Relative Centrifugal Field (× g)			k Factor*
	At $r_{max}$ (48.5 mm)	At $r_{av}$ (37.2 mm)	At $r_{min}$ (26.0 mm)	
110 000	657 000	504 000	353 000	13
105 000	599 000	459 000	321 000	14
100 000	543 000	417 000	291 000	16
95 000	490 000	376 000	263 000	17
90 000	440 000	338 000	236 000	19
85 000	393 000	301 000	210 000	22
80 000	348 000	267 000	186 000	25
75 000	306 000	234 000	164 000	28
70 000	266 000	204 000	143 000	32
65 000	230 000	176 000	123 000	37
60 000	196 000	150 000	105 000	44
55 000	164 000	126 000	88 100	52
50 000	136 000	104 000	72 800	63
45 000	110 000	84 400	59 000	78
40 000	86 900	66 700	46 600	99
35 000	66 600	51 000	35 700	129
30 000	48 900	37 500	26 200	175
25 000	34 000	26 000	18 200	253
20 000	21 700	16 700	11 700	395

\*Calculated for all Beckman Coulter preparative rotors as a measure of the rotor's relative efficiency in pelleting sample in water at 20°C.

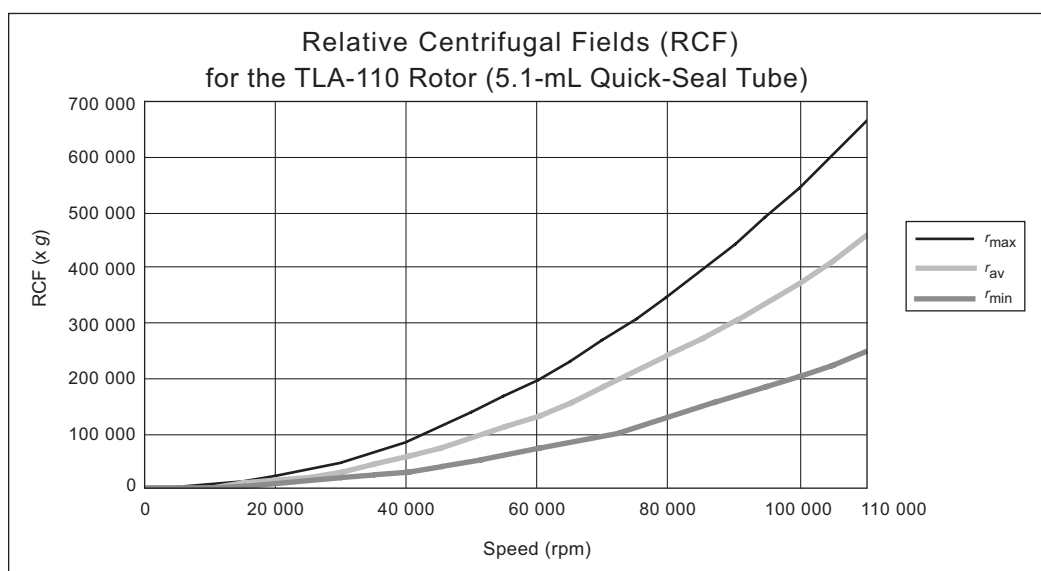


*Table 3. Relative Centrifugal Fields for the TLA-110 Rotor  
when Using 5.1-mL Quick-Seal Tubes.*

*Entries in this table are calculated from the formula  
 $RCF = 1.12r (RPM/1000)^2$  and then rounded to three significant digits.*

Rotor Speed (rpm)	Relative Centrifugal Field ( $\times g$ )			k Factor*
	At $r_{\max}$ (48.6 mm)	At $r_{av}$ (33.6 mm)	At $r_{\min}$ (18.4 mm)	
110 000	658 000	455 000	249 000	20
105 000	600 000	414 000	227 000	22
100 000	544 000	376 000	206 000	25
95 000	491 000	339 000	186 000	27
90 000	441 000	304 000	167 000	30
85 000	393 000	272 000	149 000	34
80 000	348 000	241 000	132 000	38
75 000	306 000	212 000	116 000	44
70 000	267 000	184 000	101 000	50
65 000	230 000	159 000	87 100	58
60 000	196 000	135 000	74 200	68
55 000	165 000	114 000	62 300	81
50 000	136 000	93 900	51 500	98
45 000	110 000	76 100	41 700	121
40 000	87 100	60 100	33 000	154
35 000	67 000	46 000	25 300	201
30 000	49 000	33 100	18 600	273
25 000	34 000	23 500	12 900	394
20 000	21 800	15 000	8 240	615

\*Calculated for all Beckman Coulter preparative rotors as a measure of the rotor's relative efficiency in pelleting sample in water at 20°C.



## SELECTING CsCl GRADIENTS



Precipitation during centrifugation would alter density distribution, and this would change the position of the sample bands. Curves in Figures 3 and 4 are provided up to the maximum rated speed of the rotor.

### NOTE

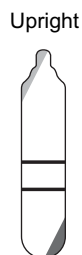
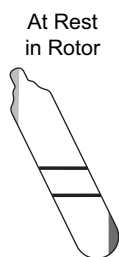
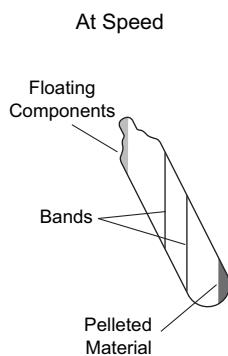
The curves in Figures 3 and 4 are for solutions of CsCl salt dissolved in distilled water only. If other salts are present in significant concentrations, the overall CsCl concentration may need to be reduced.

Rotor speed is used to control the slope of a CsCl density gradient, and must be limited so that CsCl precipitation is avoided. Speed and density combinations that intersect on or below the curves in Figure 3 ensure that CsCl will not precipitate during centrifugation in the TLA-110 rotor. Curves are provided at two temperatures: 20°C (black curves) and 4°C (gray curves).

The reference curves in Figure 4 show gradient distribution at equilibrium. Each curve in Figure 4 is within the density limits allowed for the TLA-110 rotor: each curve was generated for a single run speed using the maximum allowable homogeneous CsCl densities (one for each fill level) that avoid precipitation at that speed. (The gradients in Figure 4 can be generated from step or linear gradients, or from homogeneous solutions. But the total amount of CsCl in solution must be equivalent to a homogeneous solution corresponding to the concentrations specified in Figure 3.) Figure 4 can also be used to approximate the banding positions of sample particles.

## TYPICAL EXAMPLES FOR DETERMINING CsCl RUN PARAMETERS

**Example A:** A separation that is done frequently is the banding of plasmic DNA in cesium chloride with ethidium bromide. The starting density of the CsCl solution is 1.55 g/mL. In this separation the covalently closed, circular plasmid bands at a density of 1.57 g/mL, while the nicked and linear species band at 1.53 g/mL. At 20°C, where will particles band?



1. In Figure 3, find the curve that corresponds to the desired run temperature (20°C) and fill volume (three-quarters full). The maximum allowable rotor speed is determined from the point where this curve intersects the homogeneous CsCl density (86 000 rpm).
2. In Figure 4, sketch in a horizontal line corresponding to each particle's buoyant density.
3. Mark the point in the figure where each particle density intersects the curve corresponding to the selected run speed and temperature.
4. Particles will band at these locations across the tube diameter at equilibrium during centrifugation.

In this example, particles will band about 36 and 38 mm from the axis of rotation, about 2 mm of centerband-to-centerband separation at the rotor's 28-degree tube angle. When the tube is held upright, there will be about 2.27 mm of centerband-to-centerband separation. This interband distance,  $d_{up}$ , can be calculated from the formula:

$$d_{up} = \frac{d_{\theta}}{\cos \theta} \quad (7)$$

where  $d_{\theta}$  is the interband distance when the tube is held at an angle,  $\theta$ , in the rotor.

**Example B: Knowing particle buoyant densities (e.g., 1.59 and 1.54 g/mL), how do you achieve good separation?**

1. In Figure 4, sketch in a horizontal line corresponding to each particle's buoyant density.
2. Select the curve at the desired temperature (4°C) and fill volume (full) that gives the best particle separation.
3. Note the run speed along the selected curve (65 000 rpm).
4. From Figure 3, select the maximum homogeneous CsCl density (in this case, 1.59 g/mL) that corresponds to the temperature and run speed established above. These parameters will provide the particle-banding pattern selected in Step 2.

In this example, particles will band at about 31 and 34 mm from the axis of rotation (about 3 mm apart). When the tube is held upright there will be about 3.4 mm of center-of-band to center-of-band separation.



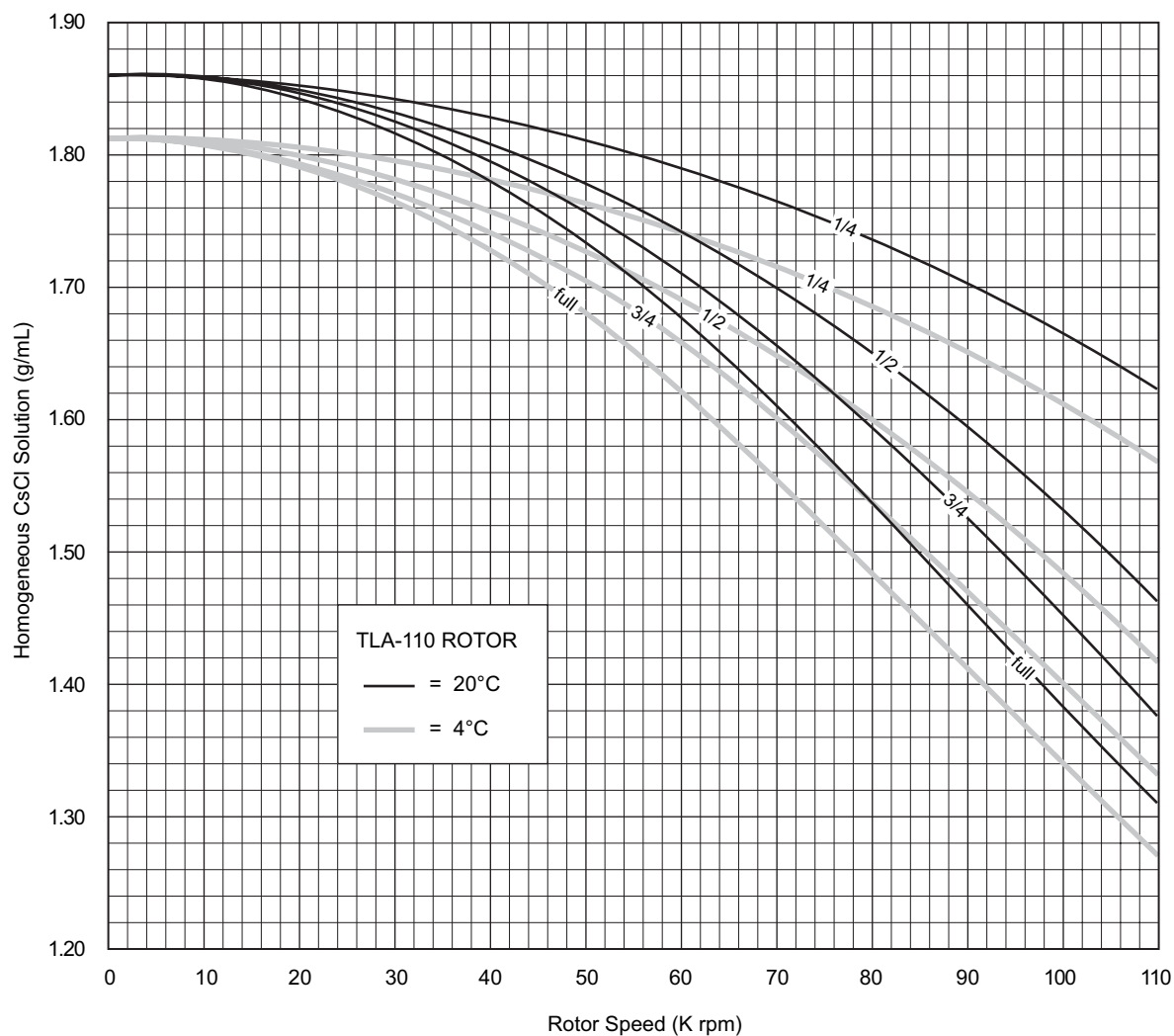


Figure 3. Precipitation Curves for the TLA-110 Rotor (3.5-mL Quick-Seal Tubes).  
Using combinations of rotor speeds and homogeneous CsCl solution densities that intersect on or below these curves ensures that CsCl will not precipitate during centrifugation.

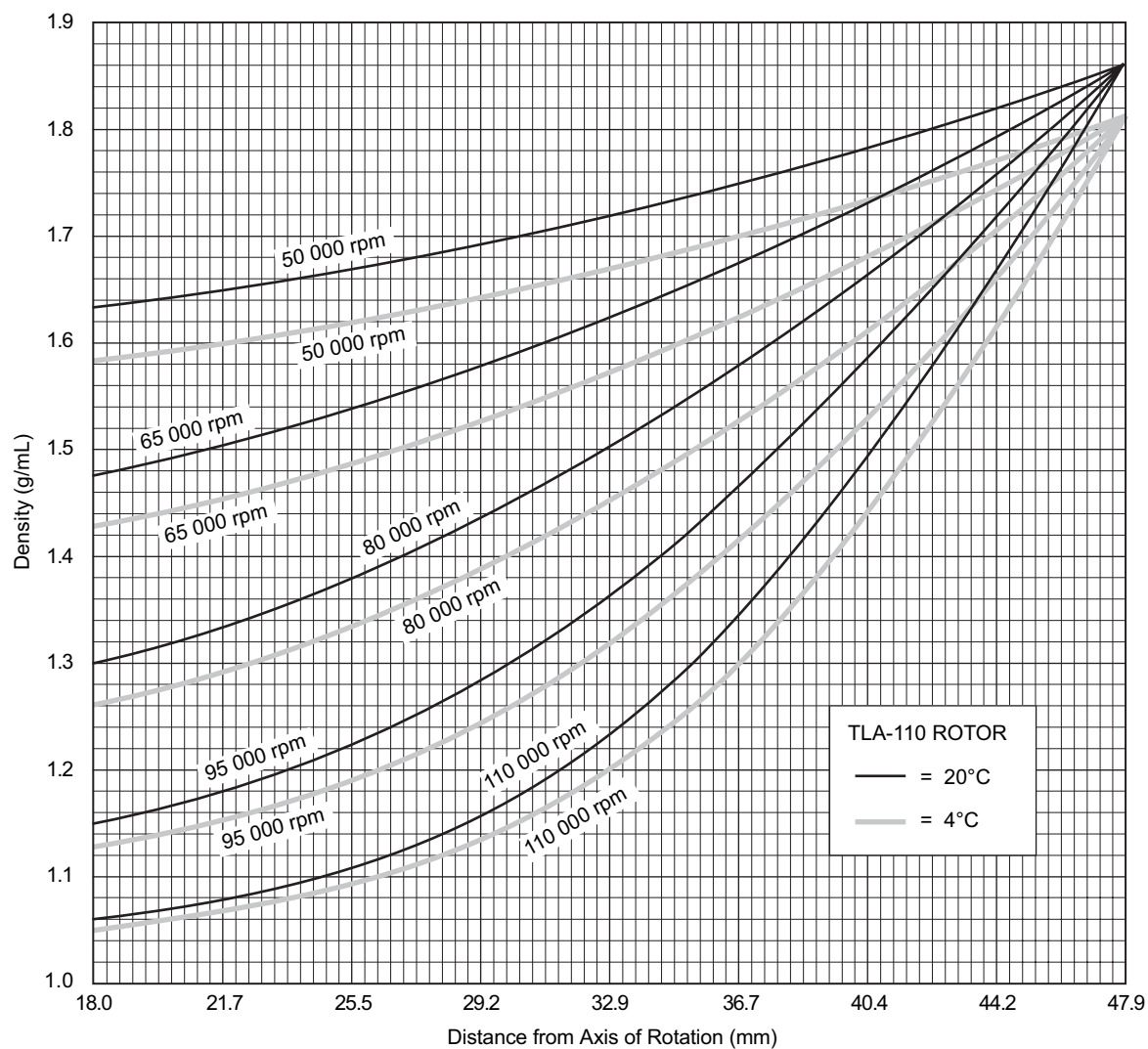


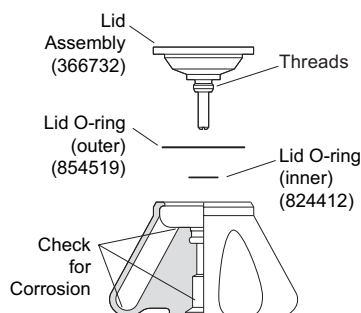
Figure 4. CsCl Gradients at Equilibrium for the TLA-110 Rotor (3.5-mL Quick-Seal Tubes).  
Centrifugation of homogeneous CsCl solutions at the maximum allowable speeds  
(from Figure 4) results in gradients presented here.

## CARE AND MAINTENANCE

### MAINTENANCE

#### NOTE

Do not use sharp tools on the rotor that could cause scratches in the rotor surface. Corrosion begins in scratches and may open fissures in the rotor with continued use.



- Regularly lubricate the metal threads in the rotor with a thin, even coat of Spinkote lubricant. Failure to keep these threads lubricated can result in damaged threads.
- Regularly apply silicone vacuum grease to the O-rings. Replace O-rings about twice a year or whenever worn or damaged.

Refer to Appendix A in *Rotors and Tubes* for the chemical resistances of rotor and accessory materials. Your Beckman Coulter representative provides contact with the Field Rotor Inspection Program and the rotor repair center.

### CLEANING



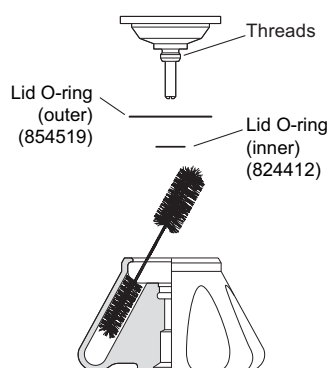
*Wash the rotor and rotor components immediately if salts or other corrosive materials are used or if spillage has occurred. Do not allow corrosive materials to dry on the rotor.*

Under normal use, wash the rotor frequently (at least weekly) to prevent buildup of residues.

1. Remove the O-rings before washing.
2. Wash the rotor and lid in a mild detergent, such as Beckman Solution 555™ (339555), that won't damage the rotor. The Rotor Cleaning Kit contains two plastic-coated brushes and two quarts of Solution 555 for use with rotors and accessories. Dilute the detergent 10 to 1 with water.

#### NOTE

Do not wash rotor components in a dishwasher. Do not soak in detergent solution for long periods, such as overnight.



3. Rinse the cleaned rotor and components with distilled water.
4. Air-dry the rotor and lid upside down. *Do not use acetone to dry the rotor.*
5. Apply a thin, even coat of silicone vacuum grease to both lid O-rings before replacing them in the grooves in the lid.

Clean metal threads as necessary (at least every 6 months). Use a brush and concentrated Solution 555. Rinse and dry thoroughly, then lubricate lightly but evenly with Spinkote to coat all threads.

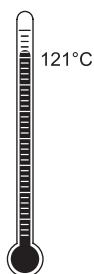
Periodically remove the O-rings and wipe clean as necessary. Clean the O-ring grooves with a cotton-tipped swab. Reapply a light film of silicone vacuum grease.

## DECONTAMINATION



If the rotor or other components are contaminated with radioactive, toxic, or pathogenic materials, follow appropriate decontamination procedures as outlined by appropriate laboratory safety guidelines and/or other regulations. Consult Appendix A in *Rotors and Tubes* to select an agent that will not damage the rotor.

## STERILIZATION AND DISINFECTION



- The rotor and all rotor components can be autoclaved at 121°C for up to an hour. Remove the lid and O-rings from the rotor and place the rotor and lid in the autoclave upside down.
- Ethanol (70%)<sup>†</sup> or hydrogen peroxide (6%) may be used on all rotor components, including those made of plastic. Bleach (sodium hypochlorite) may be used, but may cause discoloration of anodized surfaces. Use the minimum immersion time for each solution, per laboratory standards.

While Beckman Coulter has tested these methods and found that they do not damage the rotor or components, no guarantee of sterility or disinfection is expressed or implied. When sterilization or disinfection is a concern, consult your laboratory safety officer regarding proper methods to use.

<sup>†</sup> Flammability hazard. Do not use in or near operating ultracentrifuges.

## STORAGE

---

When it is not in use, store the rotor in a dry environment (not in the instrument) with the lid removed to allow air circulation so moisture will not collect in the tube cavities.

## RETURNING A ROTOR

Before returning a rotor or accessory for any reason, prior permission (a Returned Goods Authorization form) must be obtained from Beckman Coulter, Inc. This RGA form may be obtained from your local Beckman Coulter sales office, and should contain the following information:

- serial number,
- history of use (approximate frequency of use),
- reason for the return,
- original purchase order number, billing number, and shipping number, if possible,
- name and phone number of the person to be notified upon receipt of the rotor or accessory at the factory,
- name and phone number of the person to be notified about repair costs, etc.

To protect our personnel, it is the customer's responsibility to ensure that all parts are free from pathogens and/or radioactivity. Sterilization and decontamination must be done before returning the parts. Smaller items (such as tubes, bottles, etc.) should be enclosed in a sealed plastic bag.

*All parts must be accompanied by a note, plainly visible on the outside of the box or bag, stating that they are safe to handle and that they are not contaminated with pathogens or radioactivity. **Failure to attach this notification will result in return or disposal of the items without review of the reported problem.***

Use the address label printed on the RGA form when mailing the rotor and/or accessories to:

Beckman Coulter, Inc.  
1050 Page Mill Road  
Palo Alto, CA 94304

Attention: Returned Goods

Customers located outside the United States should contact their local Beckman Coulter office.

## SUPPLY LIST

Contact Beckman Coulter Sales (1-800-742-2345 in the United States; worldwide offices are listed on the back cover of this manual) or see the Beckman Coulter *Ultracentrifuge Rotors, Tubes & Accessories* catalog (BR-8101) for detailed information on ordering parts and supplies. For your convenience, a partial list is given below.

### NOTE

Publications referenced in this manual can be obtained by calling Beckman Coulter at 1-800-742-2345 in the United States, or by contacting your local Beckman Coulter office.

## REPLACEMENT ROTOR PARTS

---

TLA-110 rotor assembly . . . . .	366730
Lid assembly . . . . .	366732
Lid O-ring (outer) . . . . .	854519
Lid O-ring (inner) . . . . .	824412
Cap and plunger assembly. . . . .	349477
Spring . . . . .	347903
Rotor vise . . . . .	346133

**OTHER**

Tubes and accessories . . . . .	see Table 1
Tube rack . . . . .	355872
Quick-Seal Cordless Tube Topper kit, 60 Hz . . . . .	358312
Quick-Seal Cordless Tube Topper kit, 50 Hz (Europe) . . . . .	358313
Quick-Seal Cordless Tube Topper kit, 50 Hz (Great Britain) . . . . .	358314
Quick-Seal Cordless Tube Topper kit, 50 Hz (Australia) . . . . .	358315
Quick-Seal Cordless Tube Topper kit, 50-Hz (Canada) . . . . .	367803
Tube Topper rack . . . . .	348122
OptiSeal tube rack assembly . . . . .	361638
Spacer removal tool (for 3.5-mL and 2.0-mL Quick-Seal tubes) . . . . .	338765
Spacer removal tool (for OptiSeal tubes) . . . . .	338765
Tube removal tool . . . . .	361668
Curved hemostat (6-in.) . . . . .	927208
Straight hemostat (6-in.) . . . . .	961519
Fraction Recovery System . . . . .	342025
Fraction Recovery System Adapter Kit for TL-series tubes . . . . .	347828
Beckman Coulter CentriTube Slicer . . . . .	347960
CentriTube Slicer replacement blades (pkg of 10) . . . . .	348299
CentriTube Slicer adapter (for 13-mm tubes) . . . . .	354526
Spinkote lubricant (2 oz) . . . . .	306812
Silicone vacuum grease (1 oz) . . . . .	335148
Rotor Cleaning Kit . . . . .	339558
Rotor cleaning brush . . . . .	347404
Beckman Solution 555 (1 qt) . . . . .	339555

# ULTRACENTRIFUGE ROTOR WARRANTY

All Beckman Coulter ultracentrifuge Fixed Angle, Vertical Tube, Near Vertical Tube, Swinging Bucket, and Airfuge rotors are warranted against defects in materials or workmanship for the time periods indicated below, subject to the Warranty Conditions stated below.

Preparative Ultracentrifuge Rotors . . . . . 5 years — No Proration

Analytical Ultracentrifuge Rotors . . . . . 5 years — No Proration

ML and TL Series Ultracentrifuge

Rotors . . . . . 5 years — No Proration

Airfuge Ultracentrifuge Rotors . . . . . 1 year — No Proration

For Zonal, Continuous Flow, Component Test, and Rock Core ultracentrifuge rotors, see separate warranty.

## Warranty Conditions (as applicable)

- 1) This warranty is valid for the time periods indicated above from the date of shipment to the original Buyer by Beckman Coulter or an authorized Beckman Coulter representative.
- 2) This warranty extends only to the original Buyer and may not be assigned or extended to a third person without written consent of Beckman Coulter.
- 3) This warranty covers the Beckman Coulter Centrifuge Systems only (including but not limited to the centrifuge, rotor, and accessories) and Beckman Coulter shall not be liable for damage to or loss of the user's sample, non-Beckman Coulter tubes, adapters, or other rotor contents.
- 4) This warranty is void if the Beckman Coulter Centrifuge System is determined by Beckman Coulter to have been operated or maintained in a manner contrary to the instructions in the operator's manual(s) for the Beckman Coulter Centrifuge System components in use. This includes but is not limited to operator misuse, abuse, or negligence regarding indicated maintenance procedures, centrifuge and rotor classification requirements, proper speed reduction for the high density of certain fluids, tubes, and tube caps, speed reduction for precipitating gradient materials, and speed reduction for high-temperature operation.
- 5) Rotor bucket sets purchased concurrently with or subsequent to the purchase of a Swinging Bucket Rotor are warranted only for a term co-extensive with that of the rotor for which the bucket sets are purchased.
- 6) This warranty does not cover the failure of a Beckman Coulter rotor in a centrifuge not of Beckman Coulter manufacture, or if the rotor is used in a Beckman Coulter centrifuge that has been modified without the written permission of Beckman Coulter, or is used with carriers, buckets, belts, or other devices not of Beckman Coulter manufacture.
- 7) Rotor parts subject to wear, including but not limited to rotor O-rings, VTi, NVT™, TLV, MLN, and TLN rotor tube cavity plugs and gaskets, tubing, tools, optical overspeed disks, bearings, seals, and lubrication are excluded from this warranty and should be frequently inspected and replaced if they become worn or damaged.
- 8) Keeping a rotor log is not mandatory, but may be desirable for maintenance of good laboratory practices.

## Repair and Replacement Policies

- 1) If a Beckman Coulter rotor is determined by Beckman Coulter to be defective, Beckman Coulter will repair or replace it, subject to the Warranty Conditions. A replacement rotor will be warranted for the time remaining on the original rotor's warranty.
- 2) If a Beckman Coulter centrifuge is damaged due to a failure of a rotor covered by this warranty, Beckman Coulter will supply free of charge (i) all centrifuge parts required for repair (except the drive unit, which will be replaced at the then current price less a credit determined by the total number of revolutions or years completed, provided that such a unit was manufactured or rebuilt by Beckman Coulter), and (ii) if the centrifuge is currently covered by a Beckman Coulter warranty or Full Service Agreement, all labor necessary for repair of the centrifuge.
- 3) If a Beckman Coulter rotor covered by this warranty is damaged due to a malfunction of a Beckman Coulter ultracentrifuge covered by an Ultracentrifuge System Service Agreement, Beckman Coulter will repair or replace the rotor free of charge.
- 4) If a Beckman Coulter rotor covered by this warranty is damaged due to a failure of a Beckman Coulter tube, bottle, tube cap, spacer, or adapter, covered under the Conditions of this Warranty, Beckman Coulter will repair or replace the rotor and repair the instrument as per the conditions in policy point (2) above, and the replacement policy.
- 5) Damage to a Beckman Coulter rotor or instrument due to the failure or malfunction of a non-Beckman Coulter tube, bottle, tube cap, spacer, or adapter is not covered under this warranty, although Beckman Coulter will assist in seeking compensation under the manufacturer's warranty.

## Disclaimer

IT IS EXPRESSLY AGREED THAT THE ABOVE WARRANTY SHALL BE IN LIEU OF ALL WARRANTIES OF FITNESS AND OF THE WARRANTY OF MERCHANTABILITY AND BECKMAN COULTER, INC. SHALL HAVE NO LIABILITY FOR SPECIAL OR CONSEQUENTIAL DAMAGES OF ANY KIND WHATSOEVER ARISING OUT OF THE MANUFACTURE, USE, SALE, HANDLING, REPAIR, MAINTENANCE, OR REPLACEMENT OF THE PRODUCT.

## Factory Rotor Inspection Service

Beckman Coulter, Inc., will provide free mechanical and metallurgical inspection in Palo Alto, California, USA, of any Beckman Coulter rotor at the request of the user. (Shipping charges to Beckman Coulter are the responsibility of the user.) Rotors will be inspected in the user's laboratory if the centrifuge in which they are used is covered by an appropriate Beckman Coulter Service Agreement. Contact your local Beckman Coulter office for details of service coverage or cost.

Before shipping, contact the nearest Beckman Coulter Sales and Service office and request a Returned Goods Authorization (RGA) form and packaging instructions. Please include the complete rotor assembly, with buckets, lid, handle, tube cavity caps, etc. A SIGNED STATEMENT THAT THE ROTOR AND ACCESSORIES ARE NON-RADIOACTIVE, NON-PATHOGENIC, NON-TOXIC, AND OTHERWISE SAFE TO SHIP AND HANDLE IS REQUIRED.



# Beckman Coulter Worldwide Biomedical Research Division Offices

## AUSTRALIA

Beckman Coulter Australia Pty Ltd  
Unit D, 24 College St.  
Gladesville, NSW 2111  
Australia  
**Telephone: (61) 2 9844-6000**  
**or toll free: 1 800 060 880**  
**Fax: (61) 2 9844-6096**  
**email: lifescienceaustralia@beckman.com**

## CANADA

Beckman Coulter (Canada) Inc.  
6755 Mississauga Road, Suite 600  
Mississauga, Ontario  
Canada L5N 7Y2  
**Telephone: (1) 905 819-1234**  
**Fax: (1) 905 819-1485**

## CZECH REPUBLIC

Beckman Coulter Prague  
Radiova 1  
102 27 Prague 10  
Czech Republic  
**Telephone: (420) 267 00 85 13**  
**Fax: (420) 267 00 83 23**

## CHINA

Beckman Coulter Inc.  
Beijing Representative Office  
Unit 2005A, 2006-2009, East Ocean Center  
Jian Guomenwai Avenue  
Beijing 100004  
China  
**Telephone: (86) 10 6515 6028**  
**Fax: (86) 10 6515 6025, 6515 6026**

## EASTERN EUROPE/ MIDDLE EAST/NORTH AFRICA

Beckman Coulter International S.A.  
22, Rue Juste-Olivier  
Case Postale 301-303  
CH-1260 Nyon, Switzerland  
**Telephone: (41) 22 365 3707**  
**Fax: (41) 22 365 0700**

## FRANCE

Beckman Coulter France S.A.  
Paris Nord II, 33/66 rue des Vanesses  
B.P. 50359  
Villepinte, France  
95942 ROISSY CDG Cedex  
**Telephone: (33) 1 49 90 90 00**  
**Fax: (33) 1 49 90 90 10**  
**e-mail: bioresearch.france@beckman.com**

## GERMANY

Beckman Coulter GmbH  
Europark Fichtenhain B-13  
47807 Krefeld  
Germany  
**Telephone: (49) 21 51 33 35**  
**Fax: (49) 21 51 33 33**  
**e-mail: bioresearch.de@beckman.com**

## HONG KONG

Beckman Coulter Hong Kong Ltd.  
12th Floor, Oxford House  
979 King's Road  
Taikoo Place, Hong Kong  
**Telephone: (852) 2814 7431**  
**Fax: (852) 2873 4511**

## INDIA

Beckman Coulter India Pvt. Ltd.  
Solitaire Corporate Park  
3rd Floor - Bldg. 11  
Andheri Ghatkopar Link Road  
Chakala, Andheri West  
Mumbai India 400 098  
**Telephone: (91) 22 3080 5101**

## ITALY

Beckman Coulter S.p.a.  
Centro Direzionale Lombardo  
Palazzo F/1, Via Roma 108  
20060 Cassina de' Pecchi  
Milano, Italy  
**Telephone: (39) 02 953921**  
**Fax: (39) 02 95392264**

## JAPAN

Beckman Coulter K.K.  
TOC Ariake West Tower  
2-5-7, Ariake, Koto-ku  
Tokyo 135-0063 Japan  
**Telephone: (81) 3 5530 8500**  
**Fax: (81) 3 5404 8436**

## MEXICO

Beckman Coulter de Mexico S.A. de C.V.  
Avenida Popocatepetl #396  
Colonia Gral. Pedro Maria Anaya  
Codigo Postal 03340  
Mexico, D.F. Mexico  
**Telephone: (001) 52-55-9183-2800**

## NETHERLANDS

Beckman Coulter Nederland B.V.  
Nijverheidsweg 21  
3641 RP-Mijdrecht  
Postbus 47  
3640 AA Mijdrecht  
The Netherlands  
**Telephone: (31) 297-230630**  
**Fax: (31) 297-288082**

## SINGAPORE

Beckman Coulter Singapore Pte. Ltd.  
116 Changi Road  
Unit #03-01/02  
Singapore 419718  
**Telephone: (65) 6339 3633**  
**Fax: (65) 6336 6303**

## SOUTH AFRICA/SUB-SAHARAN AFRICA

Beckman Coulter  
Stand 1A Primegro Park  
Tonetti Street  
1685 Halfway House  
Johannesburg  
Republic of South Africa  
**Telephone: (27) 11-805-2014/5**  
**Fax: (27) 11-805-4120**  
**e-mail: beckman@intekom.co.za**

## SPAIN

Beckman Coulter España S.A.  
C/ Caleruega, 81  
28033 Madrid, Spain  
**Telephone: (34) 91 3836080**  
**Fax: (34) 91 3836096**  
**email: bioresearch.espana@beckman.com**

## SWEDEN

Beckman Coulter AB  
Archimedesvaegen 7  
Box 111 56  
SE-168 11 Bromma  
Sweden  
**Telephone: (46) 8 564 85 900**  
**Telefax: (46) 8 564 85 901**

## SWITZERLAND

Beckman Coulter International S.A.  
22, Rue Juste-Olivier  
Case Postale 301-303  
CH-1260 Nyon  
Switzerland  
**Telephone: (41) 0800 850 810**  
**Fax: (41) 0848 850 810**

## TAIWAN

Beckman Coulter Taiwan Inc.  
Taiwan Branch  
8th Floor  
216 Tun Hwa South Road, Section 2  
Taipei 106, Taiwan  
Republic of China  
**Telephone: (886) 2 2378 3456**  
**Fax: (886) 2 2377 0408**

## TURKEY

Beckman Coulter Ltd.  
E-5 Yanyol Faith Cad.  
81410 Soganlik Kartal  
Istanbul  
Turkey  
**Telephone: (90) 216 309 1900**  
**Fax: (90) 216 309 0090**

## UNITED KINGDOM

Beckman Coulter United Kingdom Ltd  
Oakley Court  
Kingsmead Business Park  
London Road  
High Wycombe  
Buckinghamshire HP11 1JU  
England, U.K.  
**Telephone: (44) 01494 441181**  
**Fax: (44) 01494 447558**  
**e-mail: beckman\_uk@beckman.com**

*Additional addresses are available at  
www.beckmancoulter.com.*



Beckman Coulter, Inc. • 4300 N. Harbor Boulevard, Box 3100 • Fullerton, California 92834-3100  
Sales and Service: 1-800-742-2345 • Internet: [www.beckmancoulter.com](http://www.beckmancoulter.com) • Telex: 678413 • Fax: 1-800-643-4366

