GeneAmp® PCR System 9700

Base Module

User's Manual



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This GeneAmp* PCR System 9700 Base Unit, Serial No_______, in combination with its immediately attached sample block modules, comprise an Authorized Thermal Cycler. The purchase price of this Base Unit includes the up-front fee component of a license under United States Patent Nos. 4,683,195, 4,683,202 and 4,965,188, owned by Roche Molecular Systems, Inc., and under corresponding claims in patents outside the United States, owned by F. Hoffmann-La Roche Ltd, covering the Polymerase Chain Reaction ("PCR") process to practice the PCR process for internal research and development using this instrument. The running royalty component of that license may be purchased from Applied Biosystems or obtained by purchasing Authorized Reagents. This instrument is also an Authorized Thermal Cycler for use with applications licenses available from Applied Biosystems. Its use with Authorized Reagents also provides a limited PCR license in accordance with the label rights accompanying such reagents. Purchase of this product does not itself convey to the purchaser a complete license or right to perform the PCR process. Further information on purchasing licenses to practice the PCR process may be obtained by contacting the Director of Licensing at Applied Biosystems, 850 Lincoln Centre Drive, Foster City, California 94404.

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Introduction and Safety

Overview

About This Chapter This chapter provides information to help you safely operate the GeneAmp® PCR System 9700.

In This Chapter The following topics are covered in this chapter:

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About This Manual

Overview

This manual describes how to use the GeneAmp® PCR System 9700. It includes the following chapters and appendixes:

- Chapter 1, "Introduction and Safety," contains safety information.
- Chapter 2, "Product Overview," describes the instrument, its components, and requirements for installation.
- Chapter 3, "Instrument Setup," provides information on how to set up the instrument and place it correctly in the laboratory, how to configure the instrument, and how to connect and configure a printer.
- Chapter 4, "Running PCR Samples," provides information on selecting a method, starting and stopping a run, reviewing the history of a run, and what to do when the run is completed.
- Chapter 5, "Creating and Editing Methods," describes how to create and edit PCR methods and how to work with stored methods.
- Chapter 6, "Converting Hold Times," provides information about setting hold times for the GeneAmp® PCR System 9700 compared to the DNA Thermal Cycler or DNA Thermal Cycler 480.
- Chapter 7, "Routine Maintenance," provides procedures for routine maintenance on the instrument.
- Chapter 8, "Troubleshooting," lists error messages and suggestions for dealing with other problems you may encounter.
- Appendix A, "Instrument Specifications," describes the dimensions, power, and electrical specifications of the GeneAmp® PCR System 9700 system, including the control panel, sample temperature information, and printer specifications.
- Appendix B, "Supplied Methods," contains information about the methods that are supplied with the instrument.
- Appendix C, "Contacting Technical Support," explains how to contact Applied Biosystems' Technical Support staff.
- Appendix D, "Limited Warranty Statement," contains the instrument warranty statement.

Instrument Safety

Instrument

Before Operating the Ensure that everyone involved with the operation of the instrument has:

- Received instruction in general safety practices for laboratories
- Received instruction in specific safety practices for the instrument
- Read and understood all related MSDSs

A CAUTION Avoid using this instrument in a manner not specified by Applied Biosystems. Although the instrument has been designed to protect the user, this protection can be impaired if the instrument is used improperly.

Documentation User Attention Words

Five user attention words appear in the text of all Applied Biosystems user documentation. Each word implies a particular level of observation or action as described below.

Note Calls attention to useful information.

IMPORTANT Indicates information that is necessary for proper instrument operation.

A CAUTION Indicates a potentially hazardous situation which, if not avoided, may result in minor or moderate injury. It may also be used to alert against unsafe practices.

A WARNING Indicates a potentially hazardous situation which, if not avoided, could result in death or serious injury.

A DANGER Indicates an imminently hazardous situation which, if not avoided, will result in death or serious injury. This signal word is to be limited to the most extreme situations.

Safety Alert Symbols

The following chart is an illustrated glossary of all electrical symbols that are used on Applied Biosystems instruments. Whenever such symbols appear on instruments, please observe appropriate safety procedures.

Electrical Symbols

	This symbol indicates the On position of the main power switch.
0	This symbol indicates the Off position of the main power switch.
Ф	This symbol indicates the On/Off position of a push-push main power switch.
<u></u>	This symbol indicates that a terminal may be connected to another instrument's signal ground reference. This is not a protected ground terminal.
	This symbol indicates that this is a protective grounding terminal that must be connected to earth ground before any other electrical connections are made to the instrument.
~	A terminal marked with this symbol either receives or delivers alternating current or voltage.
~	A terminal marked with this symbol can receive or supply an alternating and a direct current or voltage.
A	This symbol indicates the presence of high voltage and warns the user to proceed with caution.
A	This symbol alerts you to consult the manual for further information and to proceed with caution.

Electrical Safety Testing

Routine safety testing of analytical instruments (e.g., high potential voltage testing) may be required by various safety agencies.



Testing should only be carried out by qualified personnel after seeking advice from the Applied Biosystems Service Department.

Nonelectrical Symbols

The following is an illustrated glossary of all nonelectrical safety alert symbols found on Applied Biosystems instruments.



This symbol illustrates a heater hazard. Proceed with caution when working around these areas to avoid being burned by hot components.



This symbol indicates that a laser is present inside the instrument.

Chemical Safety

Chemical Hazard Warning

A WARNING CHEMICAL HAZARD. Some of the chemicals used with Applied Biosystems instruments and protocols are potentially hazardous and can cause injury, illness, or death.

- Read and understand the material safety data sheets (MSDSs) provided by the chemical manufacturer before you store, handle, or work with any chemicals or hazardous materials.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (e.g., safety glasses, gloves, or protective clothing). For additional safety guidelines, consult the MSDS.
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with adequate ventilation (e.g., fume hood). For additional safety guidelines, consult the MSDS.
- Check regularly for chemical leaks or spills. If a leak or spill occurs, follow the manufacturer's cleanup procedures as recommended on the MSDS.
- Comply with all local, state/provincial, or national laws and regulations related to chemical storage, handling, and disposal.

Chemical Waste Hazard Warning

A WARNING CHEMICAL WASTE HAZARD. Wastes produced by Applied Biosystems instruments are potentially hazardous and can cause injury, illness, or death.

- Read and understand the material safety data sheets (MSDSs) provided by the manufacturers of the chemicals in the waste container before you store, handle, or dispose of chemical waste.
- Handle chemical wastes in a fume hood.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (e.g., safety glasses, gloves, or protective clothing). For additional safety guidelines, consult the MSDS.
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with adequate ventilation (e.g., fume hood). For additional safety guidelines, consult the MSDS.
- After emptying the waste container, seal it with the cap provided.
- Dispose of the contents of the waste tray and waste bottle in accordance with good laboratory practices and local, state/provincial, or national environmental and health regulations.

About MSDSs Some of the chemicals used with this instrument may be listed as hazardous by their manufacturer. When hazards exist, warnings are prominently displayed on the labels of all chemicals.

> Chemical manufacturers supply a current MSDS before or with shipments of hazardous chemicals to new customers and with the first shipment of a hazardous chemical after an MSDS update. MSDSs provide you with the safety information you need to store, handle, transport and dispose of the chemicals safely.

We strongly recommend that you replace the appropriate MSDS in your files each time you receive a new MSDS packaged with a hazardous chemical.

A WARNING CHEMICAL HAZARD. Be sure to familiarize yourself with the MSDSs before using reagents or solvents.

Ordering MSDSs

You can order free additional copies of MSDSs for chemicals manufactured or distributed by Applied Biosystems using the contact information below.

To order documents by automated telephone service:

1	From the U.S. or Canada, dial 1.800.487.6809 , or from outside the U.S. and Canada, dial 1.858.712.0317 .	
2	Follow the voice instructions to order documents (for delivery by fax).	
	Note There is a limit of five documents per fax request.	

To order documents by telephone:

In the U.S.	Dial 1.800.345.5224, and press 1.	
	◆ To order in English, dial 1.800.668.6913 and press 1, then 2, then 1	
In Canada	◆ To order in French, dial 1.800.668.6913 and press 2, then 2, then 1	
From any other country	See the specific region under "To Contact Technical Support by Telephone or Fax (Outside North America)".	

To view, download, or order documents through the Applied Biosystems web site:

Step	Action
1	Go to http://www.appliedbiosystems.com
2	Click SERVICES & SUPPORT at the top of the page, click Documents on Demand , then click MSDS .
3	Click MSDS Index , search through the list for the chemical of interest to you, then click on the MSDS document number for that chemical to open a pdf of the MSDS.

For chemicals not manufactured or distributed by Applied Biosystems, call the chemical manufacturer.

About Waste As the generator of potentially hazardous waste, it is your responsibility to perform the Disposal actions listed below.

- Characterize (by analysis if necessary) the waste generated by the particular applications, reagents, and substrates used in your laboratory.
- Ensure the health and safety of all personnel in your laboratory.
- Ensure that the instrument waste is stored, transferred, transported, and disposed of according to all local, state/provincial, or national regulations.

Note Radioactive or biohazardous materials may require special handling, and disposal limitations may apply.

GeneAmp PCR System 9700 Labels and Warnings

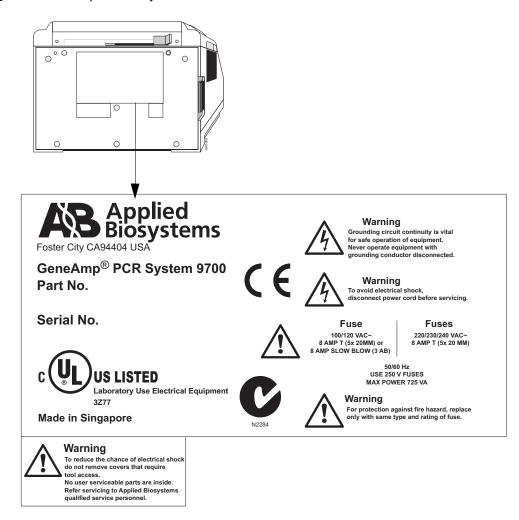
Labels

Instrument Safety Safety labels are located on the instrument. Each safety label has three parts:

- A signal word panel, which implies a particular level of observation or action (e.g., CAUTION or WARNING). If a safety label encompasses multiple hazards, the signal word corresponding to the greatest hazard is used.
- A message panel, which explains the hazard and any user action required.
- A safety alert symbol, which indicates a potential personal safety hazard.

Instrument **Warnings Diagram**

The following diagram shows where the hazards and warnings labels are located on the GeneAmp® PCR System 9700.



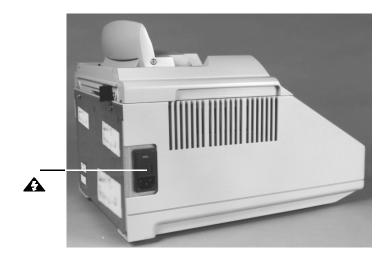
Danger of Burns

A WARNING PHYSICAL INJURY HAZARD. Hot Surface. Use care when working around this area to avoid being burned by hot components.



Hazard

Electrical Shock A WARNING ELECTRICAL HAZARD. To reduce the chance of electrical shock do not remove covers that require tool access. No user serviceable parts are inside. Refer servicing to Applied Biosystems qualified service personnel.



The three-pronged power cord and receptacle contain the grounding connector.

A WARNING ELECTRICAL HAZARD. Grounding circuit continuity is vital for safe operation of equipment. Never operate equipment with grounding conductor disconnected.



A WARNING ELECTRICAL SHOCK HAZARD. Severe electrical shock, which could cause physical injury or death, can result from working on an instrument when the high voltage power supply is operating. To avoid electrical shock, disconnect the power supply to the instrument, unplug the power cord, and wait at least 1 minute before working on the instrument.



A WARNING FIRE HAZARD. For continued protection against the risk of fire, replace fuses only with Listed and Certified fuses of the same type and rating as those currently in the instrument.



Grounding and **Electrical Safety**

The system 9700 must be grounded for protection against electrical shock.

A CAUTION ELECTRICAL HAZARD. Do not use an adapter to a two-terminal outlet since this does not provide positive ground protection.

Fuses Improper fuses can damage the wiring system and cause a fire.

A WARNING ELECTRICAL FIRE HAZARD. Before turning on the instrument, verify that the fuses are properly installed.

Laboratory Environmental Requirements

Introduction

Take the precautions described in this section whenever you operate the system 9700. Read this section before you install the instrument.

A CAUTION The instrument should be used according to the instructions provided in this manual. If used otherwise, the protection provided by this instrument may be impaired.

Temperature, Humidity, and **Environment**

IMPORTANT This instrument is designed for indoor use.

IMPORTANT Do not operate in a Cold Room or a refrigerated area. The system 9700 will operate safely when the ambient temperature is 5 °C to 40 °C (41 °F to 104 °F) and will meet performance specifications when the ambient temperature is 15 °C to 30 °C and the ambient relative humidity is 20 to 80%. These specifications have been calculated for altitudes between 0 and 2,000 meters.

A CAUTION FIRE HAZARD. This instrument is not designed for operation in an explosive environment. Do not place the instrument close to potentially explosive materials or objects.

IMPORTANT The instrument should be stored between -20 °C and 60 °C (-4 °F and 140 °F) at altitudes between 0 and 12,000 meters.

Note This instrument is able to withstand transient overvoltage according to Installation Category II as defined in IEC 1010-1.

Pollution

The installation category (overvoltage category) for this instrument is II, and it is classified as portable equipment. The instrument has a pollution degree rating of 2 and may be installed in an environment that has non-conductive pollutants only.

Emission/Immunity Statement

For our European customers, any product marked with the CE label meets the European EMC directive 89/336/EEC and the Low Voltage Directive 72/23/EEC. This product meets Class B emission limits.

Maintenance for Safe Operation

If you use any cleaning or decontamination method, except those recommended in the manual, you may risk damaging the equipment.

Maintain your instrument in good working order. In the event that the instrument has been subjected to adverse environmental conditions (such as fire, flood, earthquake, etc.), contact your local sales office for advice.

Product Overview

Overview

About This Chapter This chapter describes the GeneAmp® PCR System 9700, its components, and requirements for installation.

In This Chapter The following topics are contained in this chapter:

Topic	See page
About the GeneAmp PCR System 9700	2-2

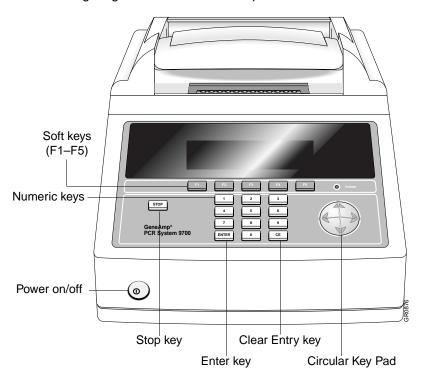
About the GeneAmp PCR System 9700

About the The GeneAmp PCR System 9700 is an automated instrument, specifically designed Instrument for the amplification of nucleic acids using the Polymerase Chain Reaction (PCR) process. The user interface consists of a control panel with a full numeric keypad, soft keys, and a graphical display screen that shows the time and temperature profile for each run.

Control Panel The instrument control panel consists of a display screen and 22 keys. The display screen shows a graphical representation of PCR events, including pre-PCR holds, PCR cycling, and post-PCR holds. You use the keys to enter information into fields on the display screen.

Diagram

Control Panel The following diagram shows the control panel.



Using the Keys The following table describes the Control Panel keys in the previous figure.

Key	Use to
Soft keys (F1-F5)	Select the function specified above the key.
	The function of each key is defined on the display screen above the key, and is redefined as you view different screens.
Numeric keys	Enter numbers from left to right into a field you highlight.
Stop key	Stop a method while it is running.
Enter key	Enter information typed into a field and advance the highlight box to the next field on a screen.
Clear Entry key (CE)	Remove information from a field.
Circular Key Pad	Move the highlight box to different fields on the display screen in the direction of the arrow.

Selecting a Field There are two ways to select a field.

If you want to	Then
move the highlight box in one of four directions	Use the Circular Key Pad.
advance the box to the next field	Press the Enter key.

Entering Numeric The following table lists how to enter numeric values for the Temperature and Hold Values Time parameters.

Parameter	Description
Temperature parameters	Enter values for temperature in decimal form. It is not necessary to type a decimal point.
	For example, to specify 89.0 °C, press 8 9 0, then press Enter.
Hold Time parameters	A hold time is the length of time the samples will be maintained at a specified temperature.
	Specify all hold times in minutes and seconds, then press Enter.
	For example, to specify one minute and five seconds, press 1 0 5, then press Enter.

Transfer or Storage You can transfer or store methods using a Methods Storage Card. The instrument can of Methods run a method from its own software or from the methods stored on a Methods Storage card.

Use	То	See
Methods Storage Card, Centennial 256kb SRAM (P/N 0940-1064)	transfer methods from the Methods Storage card to the instrument or from the instrument to the card for storage.	"Copying Methods" on page 3-25.

Firmware •

Upgrading the There are two ways to upgrade the firmware:

- With a serial cable connection (PC communication cable P/N N805-1327 or Macintosh communication cable P/N N805-1328) from the instrument's RS485 ports
- ♦ With a PCMCIA Flash Memory Card

Note Upgrade firmware can be ordered from the Applied Biosystems web site or by contacting PCR Technical Support.

For more information about upgrading the firmware, see the following.

If you are using	See
a serial cable connection (RS485 ports)	"Upgrading Through the Serial Port" on page 3-19.
a PCMCIA Flash Memory Card	"Upgrading Firmware from a PCMCIA Flash Memory Card" on page 3-23.
Microsoft Windows® 95 or Windows NT®	"Downloading Firmware Using Windows 95 or Windows NT" on page 3-21.
Windows® 3.1	"Downloading Firmware Using Windows 3.1" on page 3-22.

from the GeneAmp PCR System 9600

How the 9700 Differs The GeneAmp PCR System 9700 differs from the GeneAmp® PCR System 9600 in the following ways:

- Reaction volumes of up to 50 µL only can be run and in the "9600 Mode", as opposed to 100 µL in the GeneAmp PCR System 9600.
- Methods are stored under a user name, allowing users to keep track of and protect their own methods.
- Hold, Cycle, and Auto programs no longer exist as menu items under the Create function. Instead of linking programs together to create a method, you create a single method on the GeneAmp PCR System 9700.
- If the allowed pause time elapses during a manual pause of a method, the method will continue running (instead of stopping).
- The last method run cannot be re-run or edited unless it is first saved.
- The history file no longer includes individual setpoint time and temperature information.
- An automated restart or incubate function is available for use after a power outage.

Instrument Setup

Overview

About This Chapter

This chapter provides information on how to set up the GeneAmp® PCR System 9700 and place it correctly in the laboratory, how to configure the instrument, and how to connect and configure a printer.

In This Chapter The following topics are covered in this chapter:

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Using the Tm Calculator	3-18
Upgrading the System Firmware	3-19
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Connecting and Configuring a Printer	3-27

Unpacking the Instrument

Inspecting the Inspect the GeneAmp® PCR System 9700, and all other supplied parts, for any Instrument damage that may have occurred during shipment. If there has been any damage during transit, notify the carrier and Applied Biosystems immediately.

> Note Save the shipping container and all packing materials in case it becomes necessary to reship the instrument.

> To order additional instruments or supplies, contact one of the regional offices listed in Appendix C, "Contacting Technical Support."

Numbers

Additional Part You can order modules, accessories, and disposables from Applied Biosystems.

To order this part	Use P/N
GeneAmp PCR System 9700 Base Module	N805-0200
60-Well 0.5 mL Sample Block Module	4309131
Auto-Lid Sample Block Module	4312904
96-Well Gold Sample Block Module	4314443
96-Well Aluminum Sample Block Module	4314445
Dual 384-Well Sample Block Module	N805-0400
Temperature Verification System	N801-0435
PC (method storage) card	940-1064
Printer cable	N805-1326
PC communication cable	N805-1327
Macintosh communication cable	N805-1328

Note See the Interchangeable Sample Block Module Users Manual for part numbers of disposable items.

Setting Up your Laboratory

Choosing a Location

To prevent vibration, place the GeneAmp PCR System 9700 on a solid, stable, level surface that allows free airflow overhead and around the sides and back. You should keep all ventilation slots in the instrument cover free of obstruction, for example from excess printer paper.

Operating Temperature

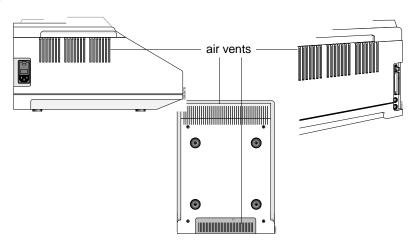
The instrument will meet performance specifications when the ambient temperature is 15 °C to 30 °C (59 ° to 86 °Fahrenheit) and the ambient relative humidity is 20% to 80%.

What to Avoid When setting up the instrument, avoid:

- Placing the instrument under overhanging shelves, especially when there is a wall behind the unit.
- Proximity to other instruments on the same bench or other heat-generating equipment.
- Locations subject to wide temperature fluctuation, such as direct sunlight, or air drafts.
- Damp areas.
- Environments where there is an oil mist.

Do Not Block Air Vents

Do not block the circulation of air to the vents located on the sides and bottom of the instrument



Note When operating multiple 9700 instruments, provide at least 8 in. of space in-between the instruments.

Configuration

Fuse Service The instrument is shipped with single-line service configuration. If your installation requires double-line service, you must remove the jumper clip. See "Converting from a Single to Double-Line Fuse" on page 3-5.

> A WARNING ELECTRICAL HAZARD. To protect yourself against shock hazards, use a properly wired three-terminal outlet. Do not use an adapter to a two-terminal outlet.

A WARNING ELECTRICAL FIRE HAZARD. Improper fuses can damage the wiring system and cause a fire. Before turning on the instrument, verify that the fuses are properly installed.

Input Voltage

IMPORTANT You must be able to disconnect the main power supply to the instrument immediately if necessary.

The following table specifies the electrical operating range for the instrument in various parts of the world. Select the appropriate fuse configuration based on the voltage used.

Location	Voltage (VAC) ^a	Frequency	Amperage (A) Nominal
Japan	100 ±10%	50/60 Hz ±1%	3.16
USA/Canada	120 ±10%	50/60 Hz ±1%	4.20
EC	230 ±10%	50/60 Hz ±1%	3.14

a. Acceptable AC line voltage tolerances: 100, 120, 220, 230 ±10%; 240 VAC +6%/-10%, 50/60 Hz ± 1%.

Note The Volt-Amp number for this instrument is 725 Volt Amps.

Modules

Interchangeable The instrument features an interchangeable sample block module allowing portability Sample Block and versatility in sample configuration.

Sample Block Module	Part Number
96-Well Gold Sample Block Module	4314443
96-Well Aluminum Sample Block Module	4314445
60-Well 0.5 mL Sample Block Module	4309131
Dual 384-Well Sample Block Module	N805-0400
Auto-Lid Sample Block Module	4312904

Note The GeneAmp PCR System 9700 will not operate without an interchangeable sample block module installed.

For an example, refer to the GeneAmp PCR System 9700 96-Well Sample Block Module User Guide, for instructions on how to install a block and configure the samples.

Converting from a Single to Double-Line Fuse

Introduction

A WARNING FIRE HAZARD. For continued protection against the risk of fire, replace fuses only with listed and certified fuses of the same type and rating as those currently in the instrument.

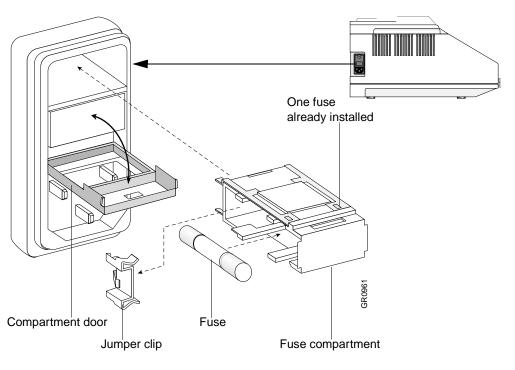
This procedure describes how to convert the instrument from the single-line configuration to double-line service (220/230/240 V nominals only).

IMPORTANT For proper operation, you must know the power source(s) available and determine if the instrument fuse configuration is correct. The instrument is shipped configured for single-line operation.

Fuse Part Number The correct fuse is an 8 amp Type T 250 V 5x20 mm fuse, P/N 0999-1683.

Power Entry Module Diagram

The following is a diagram of the Power Entry Module.



Converting Single to Double-line Fuse

The following procedure describes how to convert from a single to a double-line fuse.

Note Remove the jumper clip from the fuse compartment to convert the instrument from single-line to double-line service.

To convert from a single to a double-line fuse:

Step	Action
1	Disconnect the power cord from the power entry module.
	AWARNING ELECTRICAL SHOCK HAZARD. Severe electrical shock, which could cause physical injury or death, can result from working on an instrument when the high voltage power supply is operating. To avoid electrical shock, disconnect the power supply to the instrument, unplug the power cord, and wait at least 1 minute before working on the instrument.
2	Insert a fine flat-tip screwdriver into the slot in the upper portion of the power entry module, and open the door.
3	Pull the fuse compartment out.
	Note Observe the orientation of the metal bars that run from the fuse compartment to the two tabs.
4	Remove the jumper clip from the end portion of the fuse compartment.
5	Insert one fuse into the left side of the compartment.
6	Place the fuse compartment back into the Power Entry Module and close the fuse compartment door.
	Press the fuse compartment door until it locks in place. Connect the instrument power cord.

Turning On the Instrument

Turning On the Instrument Instrument

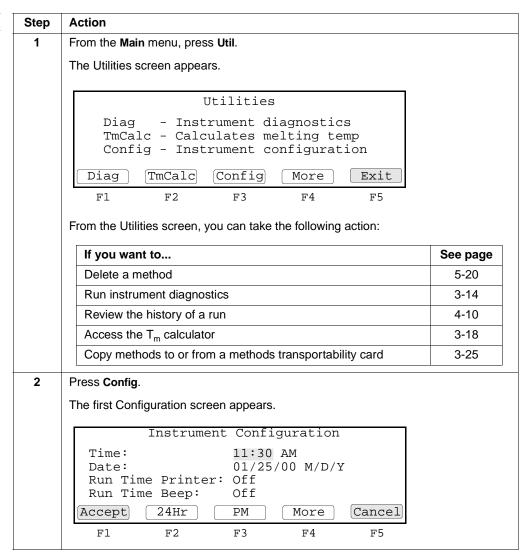
Step	Action
1	Plug the power cord into the side of the instrument and into an outlet.
2	Press the power switch to the ON position.
	The cooling fan powers up, and the Start-up screens appear.
	APPLIED BIOSYSTEMS
	www.appliedbiosystems.com
	F1 F2 F3 F4 F5
	Applied Biosystems GeneAmp® PCR System 9700 Copyright © 1996
	F1 F2 F3 F4 F5
3	After several seconds the Main menu appears. You can use any of the functions displayed above the soft keys.
	08:00 AM 4/25/01 25.0°C
	GeneAmp® PCR System 9700
	Name:tc001 User:< <pe>>></pe>
	Run Create Edit Util User
	F1 F2 F3 F4 F5
	Note The Main menu should appear within a few seconds. If any permanent patterns of lines or bars display on the screen, contact Applied Biosystems Technical Support.

Setting Custom Parameters

The instrument is shipped with default configuration values. The following procedure describes how to set customized values as well as how to enable or disable optional features.

Configuration Screen

Displaying the To display the Configuration Screen:



Setting the Time

In the first Configuration screen, you can set the current time and date for file memory maintenance, and run-time displays. You can also enable or disable the run-time printer and the run-time beep.

To set the time:

Step	Action	
1	Use the Circular Key Pad to select the Time field.	
2	Press the 24 Hr or PM (AM) soft keys until the format you want for the current time displays in the Time field.	
3	Use the numeric keys to type in the hours followed by minutes.	
4	Press Accept when your entries are complete.	
	Note CE clears an entry.	

Setting the Date You must set the instrument to the correct date.

To set the date:

Step	Action
1	Use the circular key to select the Date field.
2	The three fields to set in the Date field are the:
	◆ Days field
	◆ Month field
	◆ Year field
3	Press the D/M/Y or Y/M/D soft keys until the format you want for the current date displays in the Date field.
4	Use the numeric keys and type in a number for each of the three fields.
	The order of these three fields depends on the format you chose in step 3.

Disabling a Printer

Enabling or Enabling the printer allows you to print method parameters or records of run time events directly from the display screen. The default value for the optional printer is Off.

Note For connection and configuration see "Connecting and Configuring a Printer" on page 3-27.

To enable or disable the optional printer:

Step	Action			
1	Select the Run Time Printer field. This changes the functions of the soft keys.			
2	You can take the following action:			
	If you want to Then			
	enable the printer	Press ON.		
	disable the printer Press Off.			
3	Press Enter to accept your entry.			
4	Press Cancel to cancel all entries and return to the previous screen.			

On or Off

Turning the When turned on, the run-time beeper beeps during a pause and once at the Run-Time Beeper completion of a run. The default value is Off.

To turn the Run Time-Beeper on or off:

Step	Action			
1	Select the Run Time Beep field. This changes the value of the soft keys.			
2	You can take the following action:			
	If you want to Then]	
	turn the beeper on Press ON.			
	turn the beeper off Press Off.			
3	Press Enter to accept your entry.			

Setting the Pause The Pause Time Out field sets the time in minutes:seconds format for the length of a Time Out pause when you use the Pause soft key to pause a run from the Run Time screen. For more information on Pausing a Run, see "Pausing a Run" on page 4-8.

To set the pause time out:

Step	Action		
1	Select More to display the second configuration screen.		
	Instrument Configuration		
	Pause Time Out: 10:00 (00:01-99:59)		
	Idle State Setpoint: 25.0°C (4.0-99.9) Baud Rate: 9600		
	Baud Rate: 9600		
	Accept More Cancel		
	F1 F2 F3 F4 F5		
2	Set the Pause Time Out field by using the numeric keys to enter a time in minutes		
	and seconds.		
	From this screen, you can also define the:		
	★ Idle state setpoint temperature.		
	·		
	♦ Baud rate for your printer port.		
3	Press Accept when all information on this screen is correct.		
	Note CE clears an entry.		

Defining the Idle State Setpoint Temperature

The Idle State Set Point temperature is the temperature at which the instrument will remain when powered up, but idle.

IMPORTANT After a run is completed or terminated, there is approximately a 30 second delay before the instrument attains the specified idle state temperature. This allows you to stop one method and start another before the instrument temperature changes.

To define the Idle State Set Point temperature:

Step	Action	
1	Select the Idle State Set Point field.	
2	Use the numeric keys and type in a temperature between 4.0° C and 99.9 °C.	
	Note The default is 25 °C.	
3	Press Enter to accept your entry.	

Rate for Your **Printer Port**

Defining the Baud The following section contains a table that lists serial board specifications and a procedure that describes how to define the printer port value in the Baud Rate field.

Serial Board Specifications

You can connect the GeneAmp PCR System 9700 to any printer with a serial interface board and the following specifications.

Baud Rate	9600
Parity	NONE
Data Bits	8
Stop Bits	1

How to Set the Baud Rates

To set baud rates:

Step	Action		
1	Select the Baud Rate field.		
2	You can take the following action:		
	If you want to	Then	
	increase the baud rate	Press Up.	
	decrease the baud rate Press Down.		
	Note Available baud rates are 19200, 9600, 4800, 1200, 600, and 300.		
3	Press Enter to accept your entry.		

Configuring the Contrast and Screen Saver Options

You can use the third Configuration screen to set:

- Screen contrast from 1 to 20 (light to dark)
- Screen saver mode

Displaying the Screen

From the second Configuration screen, choose More to display the third Configuration screen.

Screen Saver Options

The following table lists the screen saver options:

Use This Option	To Activate the Screen Saver
Smart	After 15 minutes unless a method is running
Always	After 15 minutes
Never	Never

IP Address

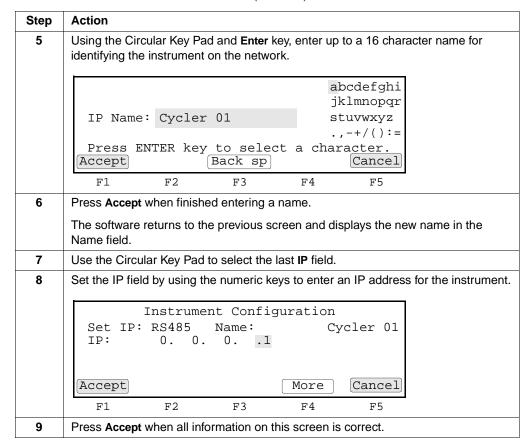
Setting the You can use the fourth Configuration screen to set the IP address for the instrument. The instrument can be linked to a network via the RS485 ports located on the right-rear side of the instrument (see the figure on page 3-19). Once connected and configured, the instrument will be visible from the network under the IP address and instrument name you select in the following procedure.

> Note See the System 9700 Networking Software User's Manual (P/N 4309575) for more information about networking GeneAmp PCR System 9700 instruments.

To set the IP Address for an instrument:

Step	Action			
1	Select More to display the second configuration screen.			
	Instrument Configuration Set IP: RS485 Name: Cycler 01 IP: 0. 0. 0. 1			
	Accept - + Cancel			
	F1	F2 F3 F4 F5		
2	Set the Set IP field using the + and – keys.			
	The following settings are available:			
	Setting Definition			
	RS485 Activates the RS 485 ports.			
	Must be used with the networking software			
	Off Turns off the networking capability and turns on the RS232 ports.			
3	Use the Circular Key Pad to select the Name field.			
4	Select Name.			

To set the IP Address for an instrument: (continued)



Running Instrument Diagnostics

Overview The instrument provides a number of internal diagnostic utilities, tests you can run to verify that the instrument hardware and software components meet performance specifications.

Diagnostic Screen

Viewing the The following procedure describes how to view the Diagnostic screen.

To view the Diagnostic screen:

Step	Action		
1	From the Main menu, press Util .		
	The Utilities screen appear	rs.	
	-	Utilities	
		rument diagnostics	
		ulates melting temp rument configuration	
	Diag TmCalc	Config More Exit	
	F1 F2	F3 F4 F5	
2	From the Utilities screen, p	oress Diag.	
	The Diagnostics screen ap	ppears.	
		agnostics	
		ware Diagnostics em Performance Tests	
	TmpVer - Temp	erature Verification	
	Upgrad - Firm	ware Upgrade	
	Hard System	TmpVer Upgrad Exit	
	F1 F2	F3 F4 F5	
3	Press the soft key that acc	esses the diagnostic utility you want to run:	
	Press this soft key	То	
	Hard	Access hardware diagnostic utilities that allow you to test hardware and electrical components such as the	
		Display screen and Keypad.	
	System	Access the system performance tests.	
		You can run these two system tests to test the rate of	
system heating and cooling, and the PCR cycling.		system heating and cooling, and the performance of PCR cycling.	
	TmpVer	Access the temperature verification tests that allow you to verify sample block calibration and uniformity.	
		These two tests are discussed in the users manual for the interchangeable sample block.	
	Upgrade	Access a utility that allows you to update the 9700 instrument firmware through the RS485 serial port or using a PCMCIA Flash Memory card.	

Electrical Components

Testing the Use the hardware diagnostic utilities to test the electrical components of the 9700 instrument.

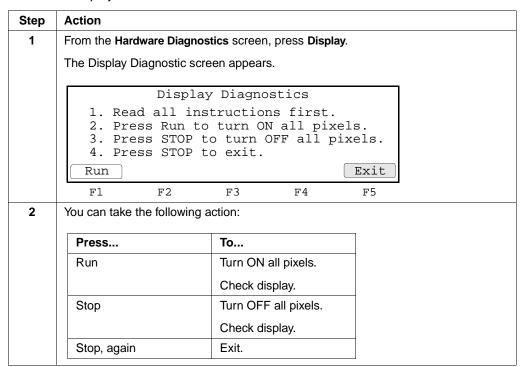
To test the electrical components:

Step	Action		
1	From the Diagnostics screen, press Hard .		
	The Hardware Diagnostics appears.		
	Hardware	Diagnostics	
	Disp - LCD Display Diagnostic Keypad - Keypad Diagnostic		
	Disp Ke	eypad Exit	
	F1 F2	F3 F4 F5	
2	Use the corresponding soft key to select the hardware component for which you want to perform one of the following tasks:		
	Press this soft key To		
	Display Visually determine if the Liquid Crystal Display (LCD) screen is functioning properly.		
	Keypad Verify the operation of the keypad.		

Screen

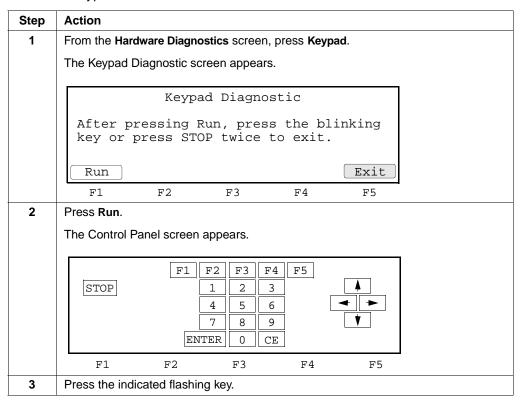
Testing the Display The Display diagnostic test allows you to visually determine if the display screen is properly functioning by turning on and off all the LCD pixels.

To test the display screen:



Testing the Keypad Use the Keypad diagnostic test to verify that all 22 keys on the control panel are functioning properly.

To test the keypad:



Using the T_m Calculator

How to Use the T_m Calculator to determine the annealing temperature of a primer set of known sequence.

To use the $T_{\rm m}$ Calculator:

Step	Action			
1	From the Main menu, press Util . The Utilities screen opens as shown below.			
	Utilities			
	Diag - Instrument diagnostics			
	TmCalc - Calculates melting temp			
	Config - Instrument configuration			
	Diag TmCalc Config More Exit			
	F1 F2 F3 F4 F5			
2	Press TmCalc.			
	The Tm Calculator appears.			
	[Salt]: 50 mM [Primer] 0.20 uM P1: 5'			
	P2: 5'			
	Tm of P1= Tm of P2=			
	Press ENTER to calculate Tm's			
3	Enter the salt concentration.			
	Note The default is 50. Enter values 5 to 1000.			
4	Enter the primer concentration.			
	Note The default it 0.20. Enter values 0.01 to 10.00.			
5	Enter primer sequence in P1.			
6	Enter primer sequence in P2 and press Enter to calculate the T _m s.			
	The melting points are displayed. Use this information to program a run.			
	For more information, see Chapter 5, "Creating and Editing Methods."			
7	Press Return to display the Utilities screen.			

Upgrading the System Firmware

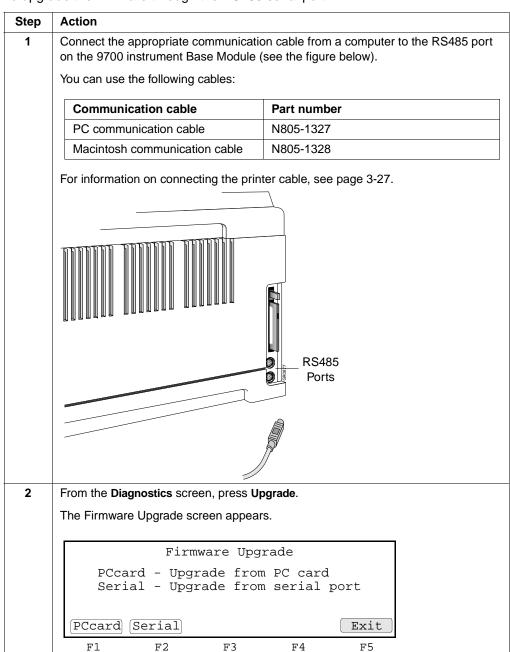
Introduction You can update the GeneAmp PCR System 9700 firmware from a serial connection to the RS485 port or from a PCMCIA Flash Memory Card.

> Contact your local Applied Biosystems representative to obtain a PCMCIA Flash Memory Card.

Upgrading Through the Serial Port

The following procedure describes how to upgrade the firmware through the RS485 serial port.

To upgrade the firmware through the RS485 serial port:



To upgrade the firmware through the RS485 serial port: (continued)

Step	Action				
3	From the Firmware Upgrade screen, press Serial. Serial Port Firmware Upgrade Attach cable from RS485 port on 9700 to				
	serial port on computer. Press Update, then begin download of file from the computer.				
		Update	Exi	Lt	
	F1 F2	F3 E	'4 F5	5	
	IMPORTANT Do not ren is completed.	nove the cable or to	urn off the instr	rument until the upgrade	
4	From the Serial Port Firmv	vare Upgrade scree	n, press Upda	te.	
	DO NOT TURN OFF THE INSTRUMENT OR REMOVE SERIAL CABLE!				
	Upgrading over the serial port				
	will take approximately 10 minutes. Instrument will re-boot when complete.				
	F1 F2 F3 F4 F5				
5	Take the following action:				
	If you are using See				
	Microsoft [™] Windows [®] 95, or Microsoft Windows NT [®] "Downloading Firmware Using Windows 95 or Windows NT" on page 3-21.				
	Microsoft Windows® 3.1	"Downloading F page 3-22.	irmware Using	Windows 3.1" on	

Firmware Using Windows 95 or Windows NT

Downloading To download the system firmware using Windows 95 or Windows NT:

Step	Action				
1	Click Start > Programs > Accessories > HyperTerminal.				
2	Click the HyperTerminal icon.				
	The Connection Description dialog box appears.				
3	In the Connection De	scription dialog box, enter the following information and click ing dialog box appears.			
	OK. The Connect Os	ing dialog box appears.			
	In this field	Take this action			
	Name	Enter a name, for example TEC.			
	Icon	Choose one of the icons.			
4	Com 2, depending or	dialog box, connect using either Direct to Com 1 or Direct to the port to which the cable is attached. NT applications this dialog box is titled Connect To , and the			
5	a. a. a com rana	er you chose Com 1 or Com 2 in the previous step, in the			
	In this field	Enter			
	Bits Per Second	9600			
	Data Bits	8			
	Parity	None			
	Stop Bits	1			
	Flow Control	None			
	When you have com	peted entering the information, click OK .			
6	From the Transfer me	enu, choose Send file .			
	The Send File dialog	box appears.			
7	In the Send File dialog box, enter the following and click Send.				
	In this field	Take this action			
	Filename	Locate the upgrade file with the name under which you saved the file.			
	Protocol	Enter Kermit.			
	A dialog box appears that displays the current packet number and the number remaining. When the download is complete, the GeneAmp PCR System 9700 automatically resets.				
8	From the File menu,	choose Save to save the parameters.			

Firmware Using Windows 3.1

 $\begin{tabular}{ll} \textbf{Downloading} & \textbf{To download the system firmware using Windows 3.1:} \\ \end{tabular}$

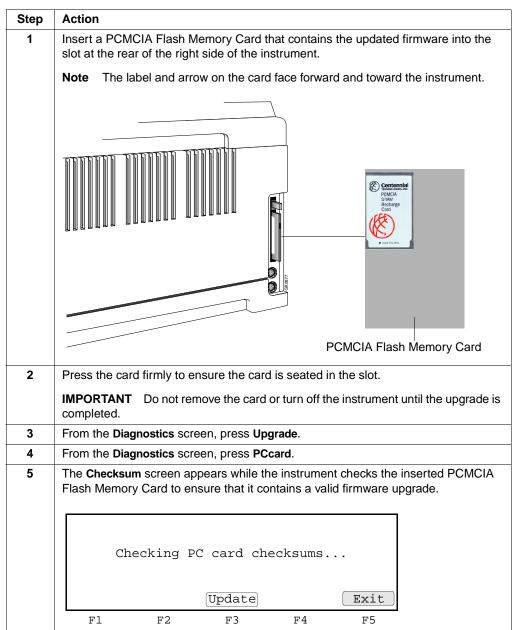
Step	Action					
1	From the Program Manager, click the Terminal program icon.					
2	From the Settings menu, choose Communication . A dialog box appears.					
3	Enter the following information in the dialog box and click OK .					
	In this field	Enter				
	Baud Rate	9600				
	Data Bits	8				
	Stop Bits	1				
	Parity	None				
	Flow Control None Connector Com1 or Com2 (depending on the port to which the cable is attached).					
4	From the Binary Tra	ansfers menu, choose Settings. A dialog box appears.				
5	Select Kermit and	click OK .				
6	From the Send Bin	ary File menu, choose Transfers.				
	A directory dialog b	pox appears.				
7	From the directory	dialog box, find and select the 9700 upgrade file , and click OK .				
	A message at the bottom of the window indicates that the program is sending the file and a progress bar appears.					
	When the download is complete the GeneAmp PCR System 9700 automatically resets.					
8	When the download is complete, from the File menu, choose Save to save these parameters.					

Upgrading Firmware from a **PCMCIA Flash** Memory Card

The following procedure describes how to upgrade the firmware using a PCMCIA Flash Memory Card.

Note Contact your local Applied Biosystems representative to obtain a PCMCIA Flash Memory Card.

To upgrade using a PCMCIA Flash Memory Card:



To upgrade using a PCMCIA Flash Memory Card: (continued)

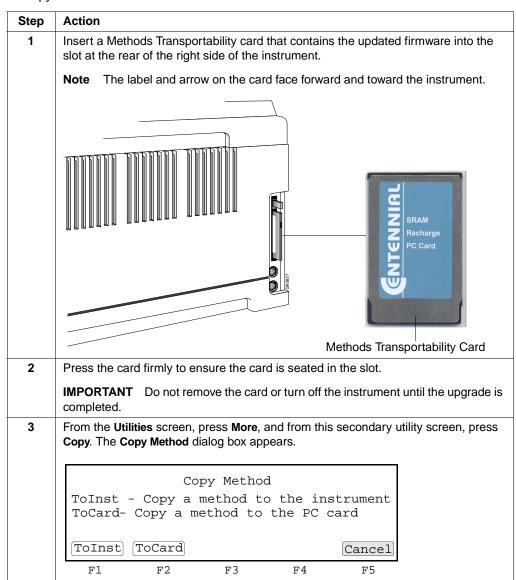
Step	Action				
6	If the PCMCIA Flash Memory Card contains a valid firmware upgrade, the Upgrade screen appears.				
	DO NOT TURN OFF THE INSTRUMENT OR REMOVE THE PC CARD!				
	Upgrading the instrument firmware will take approximately 30 seconds. Instrument will re-boot when complete.				
	F1 F2 F3 F4 F5				
	If successful, the Firmware Upgrade Successful screen displays the firmware version to which you have upgraded. Pressing the Run key in the lower right corner brings up the Main menu with the new revision number of the software listed on the screen.				
7	Remove the PCMCIA Flash Memory Card after you have successfully upgraded the firmware.				

Copying Methods

Methods

How to Copy You can copy methods from a Methods Transportability card to the instrument or from the instrument to a Methods Transportability card (P/N 940-1064). Runs can be made directly from methods on a Methods Transportability card.

To copy a method:



To copy a method: (continued)

Step	Action					
4	In the Copy Method dialog box, you can take the following action:					
	Press	То				
	Tolnst	Transfer a		m a Methods	Transportat	oility card to the
	ToCard		a method fro ability card.	m the instrun	nent to a Me	thods
	A screen appears with a list of the methods on the instrument or the Methods Transportability card, depending on whether you selected Tolnst or ToCard.					
	Methods XL PCR Touchdow AmpliTaq AmpliCyc	n PCR Gold™	Use < <pe <<pe <<pe< th=""><th>e>> 11 e>> 6 e>> 9</th><th>Stored 9/9/96 9/9/96 9/9/96 9/9/96</th><th></th></pe<></pe </pe 	e>> 11 e>> 6 e>> 9	Stored 9/9/96 9/9/96 9/9/96 9/9/96	
	Сору	[View]	User	Sort	Cancel	
	F1	F2	F3	F4	F5	
5	Use the Circu			-		
6	Confirmation	appears whe	n the metho	d has been s	successfully	copied.
		Co	opy Metho	od]
	Me		XX was su to the P	ıccessful C card.	ly	
	Copy and Yes	other met	hod?			
	F1	F2	F3	F4	F5	

Connecting and Configuring a Printer

Connecting a Printer If you connect an optional printer to your 9700 instrument, you can print out a hard copy of the time and temperature parameters for the PCR methods you create.

> Connect one end of your printer cable (N805-1326) to the RS-485 serial port on the side panel of the 9700 instrument and connect the other end to the RS-232C interface serial adapter on the rear panel of the printer.

Printer

Configuring the After you have connected the printer cable to the printer, you must configure the instrument for the printer. You can connect the 9700 instrument to any printer with a serial board and the following specifications:

Baud Rate	9600	
Parity	NONE	
Data Bits	8	
Stop Bits	1	

See your printer manual for instructions on how to complete any other necessary installation steps.

Running PCR Samples

Overview

About This Chapter This chapter provides information on selecting a method, starting and stopping a run, reviewing the history of a run, and what to do when the run is completed.

In This Chapter The following topics are covered in this chapter:

Topic	See page
Selecting a Method	4-2
Running a Method	4-6
Reviewing the History of a Run	4-10
When a Run Completes	4-12

Note For information on loading and unloading samples, see the instructions accompanying your interchangeable sample block module.

Selecting a Method

Introduction After you have prepared your samples and loaded them in the sample block, you can run a PCR amplification with a new or a stored method.

What Is a Method A method is a set of instructions in which you specify how the instrument should heat and cool your samples in a PCR thermal profile.

Methods are stored in the instrument software.

Predefined Methods The GeneAmp® PCR System 9700 supplies eight predefined methods that you can

- AmpliCycle® Sequencing
- AmpliTaq Gold®
- BigDye™
- General PCR
- LSM2
- Time Release PCR
- Touchdown PCR
- XL PCR

Each of these methods is stored under the user name <<pe>>>. You can edit these methods and store them under a different name, a different user name, or select any one and run it.

For a detailed description of each of these pre-coded methods, and how you can use them, see Appendix B, "Supplied Methods."

Selecting a Method If the method you want to run has already been created and stored, you can select it from a list. If the method you want to run has not been created, see Chapter 5, "Creating and Editing Methods."

To select a method:

Step	Action				
1	From the Main menu, press Run.				
	Note If a PC card is inserted, choose whether to run the method from the PC card or the instrument.				
	The Stored Methods screen appears.				
	Methods on Inst User Size Stored				
	exp001 lisa 10 06/23/96 exp002 lisa 15 06/25/96				
	Start View User Sort Cancel				
	F1 F2 F3 F4 F5				
	Note Stored represents the date the method was last saved. In the appropriate case, this column designates the date last used.				
2	The units for the Size field are based on a calculation of the complexity and length of a method relative to a maximum size of 1102 size segments for the storage capacity of the instrument.				
	If you need help deciding which method to select you can:				
	♦ View method parameters.				
	♦ Sort methods by different categories.				
	♦ Search for a method by user name.				
3	Select a method by using the Circular Key Pad to move the highlight box to a method listed on the Stored Methods screen.				
	Note You can use the up and down keys as repeat keys for quick scrolling.				
4	The top line of the display continuously cycles between the following three lines:				
	♦ Methods on Inst User Size Stored.				
	♦ Used Mem: xxx methods xxx segments.				
	Note The Used Mem field displays the number of size segments used by all stored methods.				
	◆ Free Mem: xxx methods xxx segments.				
	Note The Free Mem field displays the number of size segments available to store created methods.				
5	Press Start and start running your samples (see "Running a Method" on page 4-6).				

Viewing Method To view parameters Parameters

Viewing Method To view parameters of a method before running:

Step	Action				
1	From the Stored Methods screen, press View.				
	The View Method screen appears. The screen shows all the parameters of the method you selected.				
	2 Hld 3 Tmp 25 Cycles 2 Holds 94.0 94.0 55.0 10:00 0:30 55.0 0:30 5:00 0:30 5:00 0:30 5:00				
	Start Method: exp 001 Return				
	F1 F2 F3 F4 F5				
2	After reviewing PCR and post-PCR parameters of a stored method, you can:				
	♦ Press Start to start the method.				
	♦ Press Return and return to the Stored Methods screen.				
	Note You cannot edit parameters from the View Method screen.				

Searching for Methods

Searching for You can find any method that has been stored under a user name.

To search for a method:

Step	Action				
1	From the Stored Methods screen, press User.				
	Select which user's methods to view by taking the following action.				
	If you want to	Then			
	list all the methods currently stored on the instrument Press All.				
	display the methods stored under that user's name				
	Note You cannot add, delete, or modify a user name from this screen.				
2	Making a selection returns you to the Stored Methods screen which now displays the methods of the user you selected.				

 $\textbf{Sorting Methods} \quad \text{If you have a large number of stored methods, you can sort them by name, date last}$ used, date stored and size.

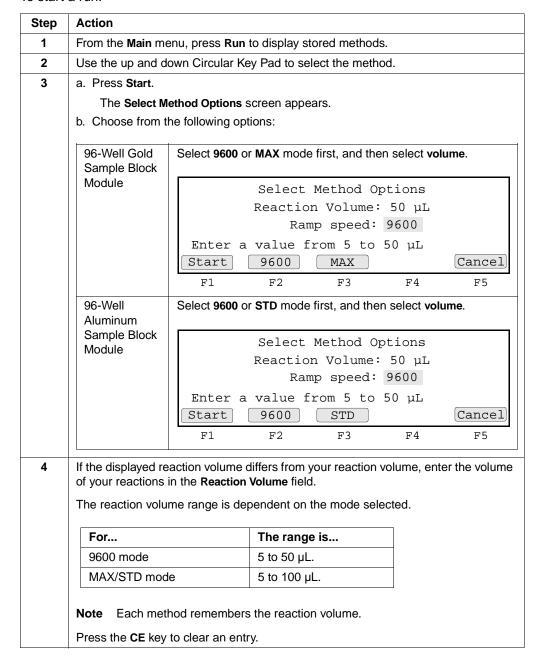
To sort methods:

Step	Action		
1	From the Stored Methods screen, press Sort.		
	The sorting criteria screen appears.		
	Sort Methods By: Method name		
	Date last u		
	Date stored Method size		
	[Accept]	Cancel	
	F1 F2 F3	F4 F5	
2	Use the up and down Circular Key Pad to	select the type of sort.	
	The following table describes the sort met	thods:	
	Choose this item	To sort	
	Method name	Methods alphabetically.	
	Date last used	Methods chronologically in descending order by date of use.	
		The last method which ran or was stored is listed first.	
	Note Uses the most recent title and	Methods chronologically by date stored.	
	date, between date last used and the data stored.	The last method stored is listed first.	
	Method size	Methods in increasing order by the amount of memory used to store each method.	
		The largest size method is listed first.	
3	Press Accept to accept a selection.		
	This returns you to the Stored Methods so sorted according to your selection in step		

Running a Method

Starting a Run After choosing a method, follow these steps to start a run.

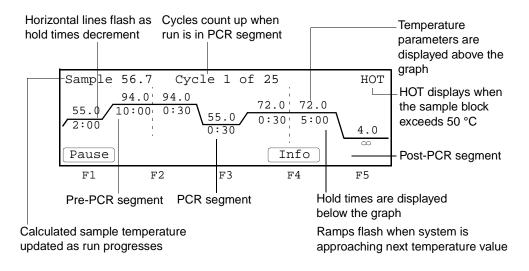
To start a run:



To start a run: (continued)

Step	Action
5	Press Start to start a run.
	If the temperature of the heated cover is less than 103 °C, this message, "Cover is heating" appears.
	Please wait. Cover is heating
	Current temperature: 65°C The run will begin when the heated cover reaches 103°C.
	Cancel
	F1 F2 F3 F4 F5
6	When the heated cover reaches 103 °C, the Run Time screen displays and the method you selected starts running.
	For a description of this screen, see "About the Run Time Screen" on page 4-7.
	Sample 56.7 Cycle 1 of 25 HOT
	$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$
	F1 F2 F3 F4 F5

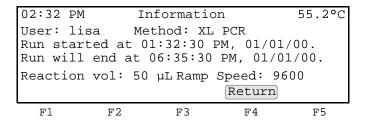
About the Run Time You can use the Run Time screen to chart progress at any time during the run. The Run Time screen displays the executing segment, and the next segment to execute.



Use the Run Time screen for	For more information, see page
Viewing Method Information	4-8
Pausing a Run	4-8
Stopping a Run Before It Completes	4-9

Information

Viewing Method You can view the Method Information screen during a run by pressing Info. Press Return to return to the Run Time screen.



Pausing a Run

You can manually pause a run for a ten minute period of time during a run by pressing Pause (Figure 4-1). If you want to specify a different period of time for a pause, see "Setting the Pause Time Out" on page 3-10.

Press Resume to resume running a method before a pause expires.

Note You can pause a run in order to add a reagent. Do not to touch the sample block or the bottom of the heated cover during a pause.

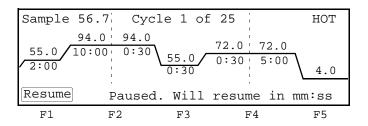


Figure 4-1 Pause screen

Your samples remain at the temperature of the instrument when you pause the run. The time remaining in a pause appears at the bottom of the screen in minutes:seconds format. It decrements to zero, and the paused run resumes at the point where you paused it.

Before It Completes

 $\begin{tabular}{ll} Stopping a Run & The following procedure describes how to stop a run before it completes. \\ \end{tabular}$

To stop a run before it completes:

Step	Action		
1	Press the Stop key.		
	The Stop confirmation screen appears.		
	Sample 50.1 Confirm Stop HOT		
	Press STOP to abort. Press Resume to continue.		
	Resume		
	F1 F2 F3 F4 F5		
	The run pauses for a pre-programmed period of time. When the pause time expires, the run aborts.		
	Press Resume to resume.		
	Note The stop and pause times are configured by setting the Pause Time Out . See "Setting the Pause Time Out" on page 3-10.		
2	Press the Stop key again.		
	This stops the run and the End of Run screen appears.		
	11:30 AM End of Run 25.1°C		
	Method:exp001 Run aborted at 11:30:05 AM 01/01/00. Length of run is 01:34:25.		
	Hist		
	F1 F2 F3 F4 F5		
	If any errors occur during a run the following message appears, "Exception occurred, check history file." Press HIST to review the history file.		
3	Press Exit to return to the Main menu.		

Reviewing the History of a Run

How to Review the You can read a record of the events and errors that occurred during a run by reviewing History of a Run the history file. The instrument stores the history file until it is overwritten by the next method used.

To review the history:

Step	Action		
1	To display the History File screen:		
	♦ From the Utilities screen, press More , and then press Hist .		
	♦ From the End of Run screen, press Hist.		
	History of method exp002 User: lisa Reaction volume: 50 µL Run started at 02:30:45 PM, 01/01/00. Run aborted at 02:50:42 PM, 01/01/00. Length of run 00:19:57 Ramp speed: 9600 No exceptions PageDn Print Return		
	F1 F2 F3 F4 F5		
2	Press PageUp to move up through the record, or PageDn to move down.		
3	Press Print to print the record.		

History Formats The following table lists the history line formats.

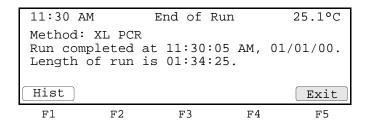
Pre-PCR hold	<exception> in Pre-PCR xx Setpt xx</exception>
PCR segment	<exception> in Cycle xx Setpt xx Repxx</exception>
Any other hold	<exception> in Hold xx Setpt xx</exception>

$\label{thm:cond} \textbf{History File Records} \quad \text{The following table lists the history file records.}$

Record	Description	Туре
History of method xxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxx	Note If you stopped a method before it completed running, then the message, Run ended at, will be Run aborted at	Report
Power failure in Cycle xx at Setpt xx. Power failed at hh:mm:ss am for hh:mm:ss. Run resumed at hh:mm:ss am	There was a power failure during a specified point in a cycle. The message, for >18, indicates that the power was off for more than 18 hours.	Report
Drift error in Cycle xx Setpt xx Repxx. Temperature drifted x.x°c from setpt	Block drift error. The block has drifted ± 2 °C from set point during the hold segment of a run.	Report
Cover error in Cycle xx Setpt xx Repxx. Heated cover at xx.x°c	Heated cover drift error. The cover has drifted ± 5 °C from 105 °C anytime during the run.	Report
Sensor error in Cycle xx Setpt xx Repxx. Block sensor failure.	Block sensor failure.	Fatal error. Call for service.
Sensor error in Cycle xx Setpt xx Repxx. Cover sensor failure.	Heated cover sensor failure.	Fatal error. Call for service.
Setpt error in Cycle xx Setpt xx Repxx. Could not reach xx.x in hh:mm:ss	This setpoint error is only logged for setpoints above 15 °C. The limit is 5 times the normal ramping time.	Fatal error. Call for service.
Program pause in Cycle xx Setpt xx Rep xxx Method paused at xx °C for hh:mm:ss	A programmed pause was encountered.	Report
Manual pause in Cycle xx Setpt xx Rep xxx Method paused at xx °C for hh:mm:ss	You paused the run.	Report

When a Run Completes

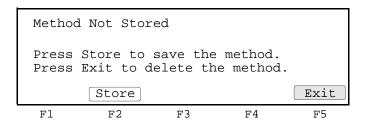
End of Run Screen At completion of a run, the instrument beeps and the End of Run screen appears. From the End of Run screen you can perform the same functions as you can from the Stop Run screen.



If you have not yet stored the method, you must store it before exiting, or you will lose the settings. The Store soft key appears if the method has not yet been stored.

Method Not Stored Screen

If you attempt to exit the End of Run screen before storing the method, the Method Not Stored screen displays.



The following table lists the actions you can take.

If you	Then press
want to store the method	Store.
do not want to store the method	Exit. Note The Method Not Stored screen appears for a few
	seconds before the Exit key is recognized. This prevents you from losing a newly created method.
	You return to the top level screen and the method you created is not saved.

Creating and Editing Methods

Overview

About This Chapter

This chapter provides information about how to create and edit PCR methods and how to work with stored methods.

In This Chapter The following topics are covered in this chapter:

Topic	See page
Adding or Changing Users	5-2
Creating Methods	5-6
Modifying Cycling	5-11
Printing a Method	5-17
Editing or Deleting Methods	5-18

Adding or Changing Users

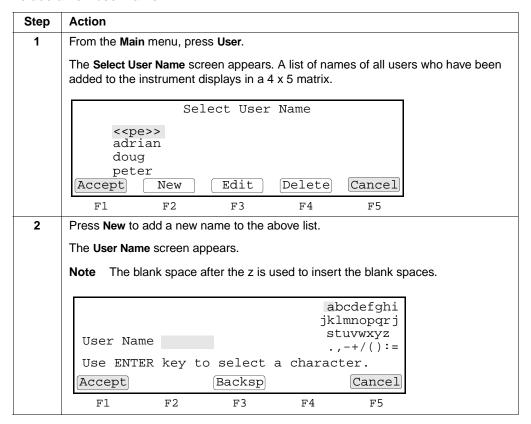
Introduction The GeneAmp® PCR System 9700 stores methods by user's names. You can add up to 19 different user names to the instrument. Once you've added your name to a list of users, and stored a method under that name, you can run the method at any time by selecting it from the Stored Methods screen (See "Selecting a Method" on page 4-3).

> Use the User function on the Main menu to add new users or edit existing user names. The name you add or the name you select from a list of existing user names becomes the current user name. All new methods that you create are stored by default under the current user name.

Name

Adding a New User You add a new user name by entering an alphanumeric name on the User Name screen.

To add a new user name:

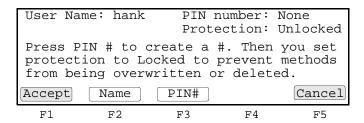


To add a new user name: (continued)

ер	Action		
3	In the User Name field, enter an alphanumeric name up to six characters in length		
	You can take the following action:		
	If you want to	Then	
	select a character in the list shown in the upper right portions of the screen	use the Circular Key Pads.	
	put the alphabetic character in the User Name field	press Enter.	
	enter the numbers directly into the User Name field	use the numeric keys.	
	go back one space and remove a	hold down the Soft key and go back	
	single character	multiple spaces.	
4	, ,	multiple spaces.	
4	Press Accept to accept a name. The Sec You can take the following action:	multiple spaces. urity Code screen appears.	
4	Single character Press Accept to accept a name. The Sec You can take the following action: If you	multiple spaces. urity Code screen appears. Then	

Protecting Methods

You can protect methods and prevent other users from accidentally overwriting or deleting them by entering a Personal Identification Number (PIN#) on the Security Code screen.



The following table lists the two levels of protection.

If a	Then other users cannot
user has entered a PIN #	edit that user's name without knowing the PIN #.
method is locked	delete/overwrite the method.

Follow the procedure below to protect a method.

To protect a method:

Step	Action		
1	Press PIN #.		
	The New PIN Number screen appears.		
	Create a PIN Number		
	Your PIN number protects the access to your user name and protection level		
	Enter a PIN number. New PIN #: XXXX		
	Accept		
	F1 F2 F3 F4 F5		
2	In the New PIN # field, use the numeric keys and type in a four-digit PIN.		
3	Press Enter.		
	The PIN Confirmation screen appears.		
	Confirm PIN Number		
	Your PIN number protects the access to		
	your user name and protection level Enter a PIN number again. PIN #: XXXX		
	Press Accept to confirm your PIN #.		
	Accept		
	F1 F2 F3 F4 F5		
4	Confirm your PIN by typing your four-digit PIN in the Confirm PIN # field.		
5	Press Enter.		
	The Protection Status screen appears.		
	Username: hank PIN number: XXXX		
	Protection: Unlocked		
	Press PIN # to create a #. Then you set protection to Locked to prevent methods		
	from being overwritten or deleted.		
	Accept Name PIN# Lock Cancel		
	F1 F2 F3 F4 F5		
6	Press Lock to lock your method.		
	This toggles between a Locked and Unlocked state. The Protection field displays the status of the method.		
7	Press Accept when you have entered a PIN you want to keep. The Select User Name screen appears.		
	The new name you entered should now display on the screen.		
	Press Cancel to cancel your entry and return to the previous screen.		

Changing a User If you know the personal identification number for a user name, you can use the Name Circular Key Pad to select that name, and change it.

To change a user name:

Step	Action
1	From the Main menu, press User.
	The Select User Name screen appears.
2	Use the Circular Key Pad to select the name you want to change.
3	Press Edit.
	If a PIN has been previously entered, the Security Check screen appears.
4	Type in the four-digit PIN of the user name you selected.
5	Press Name to enter a new user name.
6	Press the CE key to clear the previous name.
7	Enter a new user name.
8	Press Accept.
	The Security Code screen appears again.

Name

Deleting a User If there aren't any methods stored under a user name, you can delete that name from the Select User Name screen.

To delete a user name:

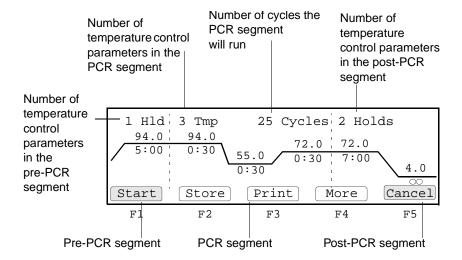
Step	Action
1	Use the Circular Key Pad from the Select User Name screen to select a user name.
2	Press Delete to delete the name.
	This removes the name from the Select User Name screen and allows you to add a new name to the instrument.

Creating Methods

Method

About the Default The GeneAmp PCR System 9700 comes with a default PCR thermal profile called a method. The create screen displays this default method. For information on displaying the create screen, see "Displaying the Create Methods Screen" on page 5-7.

> You can run the default method shown above, or use it as a template to create a customized method.



Basic Parameters

To create a method, you need to define the following four basic parameters:

- **Temperature Control Parameters**
- Pre-PCR Holds
- **PCR Parameters**
- Post-PCR Holds

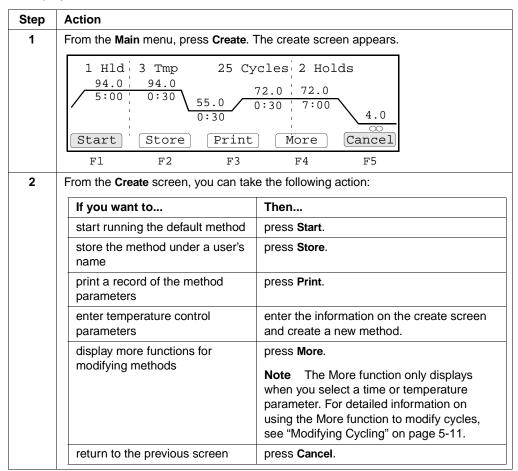
For detailed information about these parameters refer to the table below:

For information about	See page
Entering Temperature Control Parameters	5-8
Defining Pre-PCR Holds	5-9
Defining PCR Parameters	5-9
Defining Post-PCR Holds	5-10

Displaying the **Create Methods** Screen

Follow the steps below to display the Create Methods screen.

To display the Create Methods screen



Temperature **Control Parameters**

Entering When you enter temperature control parameters, you define values for parameters in each of the three segments of a method: pre-PCR, PCR, and post-PCR.

To enter temperature control parameters:

Step	Action
1	On the Create screen, select a field.
	When you first display the Create screen, the HId field is highlighted.
2	Use the numeric keys to enter values.
3	Press Enter to accept a value. The next field is then selected in the order shown in Figure 5-1.

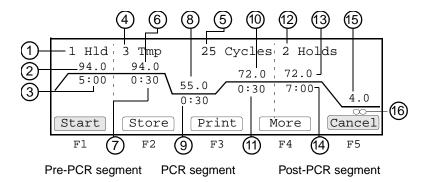


Figure 5-1 Order of advancement of the highlight box

Number	Description
1	Number of pre- PCR holds
2	Pre-PCR temperature parameter
3	Pre-PCR hold time parameter
4	Number of PCR segment temperatures
5	Number of cycles for the PCR segment
6	PCR segment temperature parameter
7	PCR segment time parameter
8	PCR segment temperature parameter
9	PCR segment time parameter
10	PCR segment temperature parameter
11	PCR segment time parameter
12	Number of post-PCR holds
13	Post-PCR temperature parameter
14	Post-PCR hold time parameter
15	Post-PCR temperature parameter
16	Post-PCR hold time parameter

Defining Pre-PCR The Hld field on the Create screen defines the number of holds for the pre-PCR Holds segment of your method. One (1) is the typical setting for most PCR amplifications and is the default value for the Hld field.

> Generally, pre-PCR holds define a temperature and hold-time sufficient to denature any endonucleases or exonucleases that may contaminate your prepared samples.

To define pre-PCR holds:

Step	Action
1	On the Create screen, in the HId field, type in the number of pre-PCR holds for your method.
	Note You can enter 0 in this field to delete the pre-PCR hold segment from your method.
2	Create the first temperature parameter:
	a. Press Enter.
	b. Type in a temperature value between 4.0 °C and 99.9 °C.
3	Create the first hold-time parameter:
	a. Press Enter.
	b. Type in a hold-time value between 00:00 and 98:59 (minutes:seconds).
4	Enter information for the next pre-PCR hold:
	a. Press Enter.
	 Repeat step 2 and step 3 until you have hold-time and temperature values for each of the pre-PCR hold parameters you defined in step 1.

Parameters

Defining PCR The Tmp field on the Create screen defines the number of temperature control parameters in the PCR cycling segment of your method. Three temperature PCR is the typical setting for many PCR amplifications:

- Template denaturation
- Primer annealing
- Primer extension

To define the PCR parameters:

Step	Action
1	On the Create screen, in the Tmp field, type in the number of temperature control parameters (2–6) you want for the PCR segment of your method.
	Note Specifying only the minimum number of PCR cycles required for analysis will minimize the chance that unwanted targets will amplify competitively.
2	Enter the number of cycles you want the method to run:
	a. Press Enter to select the Cycles field.
	b. In the Cycles field, type in the number of cycles (from 2–99).
	Note Twenty-five cycles is the default setting.
3	Create the first temperature parameter:
	a. Press Enter.
	b. Type in a temperature value between 4.0 °C and 99.9 °C.

To define the PCR parameters: (continued)

Step	Action
4	Create the first hold-time parameter:
	a. Press Enter.
	b. Type in a hold-time value between 00:00 and 98:59 (minutes:seconds).
5	Repeat step 3 and step 4 until you have time and temperature values for each of the segment temperature control parameters you defined in step 1.

Defining Post-PCR On the Create screen, the Holds field defines the number of temperature control $Holds \;\;$ parameters in the post-PCR segment of your method.

> The post-PCR incubation temperature and hold time parameters define how to hold your samples at a specified temperature until you are ready to analyze them.

Note If the idle state setpoint, or the last hold of the Method are below 15 °C, then the heated cover will automatically set to 50 °C.

Post-PCR Parameter Settings

Typical Post-PCR parameter settings:

Temperature	Time (min:sec)	Use For
72 °C	7:00	Complete extension of all amplicons
72 °C	99:59 (x)	AmpErase™ applications
4 °C	99:59 (x)	General storage

Follow the procedure below to define Post-PCR Holds

To define post-PCR holds:

Step	Action
1	Select the Holds field.
2	In the Holds field, type in the number of post-PCR steps for your method.
3	Press Enter to select the first post-PCR temperature parameter.
4	Type in a temperature value between 4.0 °C and 99.9 °C.
5	Press Enter to select the first post-PCR hold time parameter.
6	Type in a hold time value between 00:00 and 98:59 (min:sec).
	Note The hold time × indicates a hold that lasts indefinitely. You can enter an × hold time, by typing a hold time value of 99:00 or greater.
7	Press Enter. This selects the next temperature parameter.
8	Repeat step 4 through step 7 until you have time and temperature values for each of the post-PCR hold parameters you defined in step 2.

Modifying Cycling

Introduction

In addition to customizing values for PCR temperature control parameters, you can use the More function on the create screen and access cycle modification functions that allow you to:

- Auto-increment/decrement time and temperature parameters.
- Modify up-ramp and down-ramp rates in the cycling segment of a method.
- Insert holds, cycles, and programmed pauses.
- Delete temperature control parameters.

The time or temperature parameter you select on the create screen, determines which modification function you can access when you press More. Different modification functions are available depending on whether you select a temperature control parameter in the pre-PCR segment, the PCR segment, or the post-PCR segment of a method.

Changing **Temperature Control Parameters**

Using the AutoX function, you can automatically increase or decrease the value for any PCR segment parameter by a fixed amount every cycle.

Note This feature is particularly useful towards the end of the amplification process since the amount of PCR product, available to be extended, increases with the number of cycles while the amount of available enzyme remains constant.

To automatically change temperature control parameters:

Step	Action
1	From the Create screen, use the Circular Key Pad to select a time or temperature parameter in the PCR segment.
2	Press More.
	The Modify screen appears.
3	Press Modify.
	The Select Modification screen appears.
4	Press AutoX. The AutoX screen appears.
	2 Pre-PCR 3 Tmp 25 Cycles 2 Holds +0.0 +0.0 +1.0 +0.00 +1.0 +0.00 -0:30 * Cancel F1 F2 F3 F4 F5
	Note If you have inserted a programmed pause, the AutoX screen displays the pause, but you cannot modify it from the AutoX screen.
5	Select the PCR time or temperature parameter that you want modified when you run your method.
	Note From the AutoX screen, you cannot modify the number of parameters in each segment or the number of cycles.

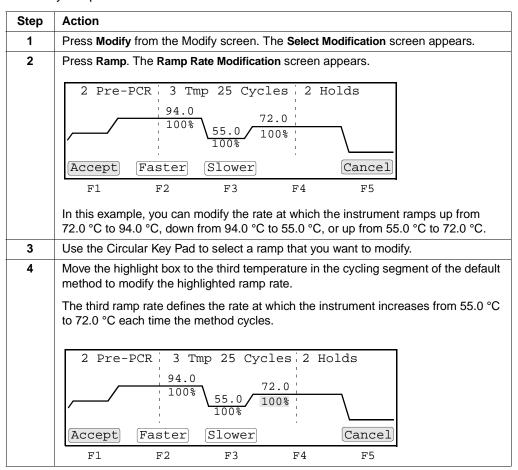
To automatically change temperature control parameters: (continued)

Step	Action		
6	You can take the following action:		
	If you want to	Then press	
	increase the value every cycle	+ (plus sign).	
	(a plus sign displays in the current field)		
	decrease the value every cycle	- (minus sign).	
	(a minus sign displays in the current field)		
	A set side * see see see see see see see see see		
	An asterisk * appears on method screens for para	imeters that have been modified	
7	Press Accept to accept all entries on the AutoX screen.		

Modifying Ramp

The ramp time is the time it takes the instrument to change from one temperature to another. Using functions accessible from the Modify screen, you can modify the up-ramp and down-ramp rates of the instrument by defining it as a percentage of the temperature's maximum rate of increase. The default maximum up-ramp and down-ramp rates is 100%.

To modify ramp rates:



To modify ramp rates: (continued)

Step	Action			
5	Modify the up-ramp and down-ramp rates you selected by defining them as a percentage of the maximum of 100%:			
	If you want to	Then press		
	increase the ramp rate by 10% up to a maximum of 100%	the Faster key.		
	decrease the ramp rate by 10% from 100% to 10%, and by 5% from 10% to 5%	the Slower key.		
	Note You can also use the numeric keys to enter a value that defines the percentage by which you want to decrease the ramp rate for each cycle of the method. You can enter values between 5 and 95, or 100.			
For ramp rates less than 100%, an asterisk * appears next to modified ran asterisk remains beneath the modified temperature parameter to remind y the method has been modified.				
	Note The following message appears if you enter a numeric va range of acceptable values, "Valid range is 5 to 95 and 100."	lue outside the		
6	Press Accept to accept all entries, and return to the previous screen	een.		

pauses that the instrument automatically inserts into your method as it runs.

To insert holds:

Step	Action	
1	From the Create screen, use the Circular Key Pad to select a time or temperature parameter to the right of where you want to insert a hold.	
2	Press More.	
	Depending on the parameter you select in step 1, one of three screens displays from which you can access the insert function.	
3	Press Insert. The Insert screen appears.	
	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	
4	Press Hold to insert a hold of 4.0 °C for 30 seconds to the left of the parameter you selected in step 1.	
5	Type in a value for the hold temperature.	
6	Type in a value for the hold time.	
7	Press More to return to the Create screen. Your modified method now appears.	

Deleting Holds Follow the procedure below to delete holds.

To delete holds:

1	From the Ed	t screen, sele	ect a hold pa	rameter and p	press More.	
2	2 Pre	-PCR 3 Tr	mp 25 Cyc	time or temp cles; 2 H 72.0; 72.0 0:30; 5:00 More	Iolds	parameter.
	F1	F2	F3	F4	F5	
3	Press Delete					_

Inserting Cycles Follow the procedure below to insert cycles.

To insert cycles:

Step	Action
1	From the Create screen, use the Circular Key Pad to select a time or temperature parameter to the right of where you want to insert a cycle.
2	Press More . Depending on the parameter you select in step 1, one of three screens displays from which you can access the insert function.
3	Press Insert. The Insert screen appears. 2 Pre-PCR 3 Tmp 25 Cycles 2 Holds 94.0 94.0 55.0 10:00 0:30 * 72.0 72.0 2:00 55.0 0:30 5:00 Hold Cycle Pause Cancel
	F1 F2 F3 F4 F5
4	Press Cycle to insert a cycle to the left of the segment you selected in step 2.
	Note You can delete a cycle by entering 0 in the Tmp field.

Programmed Pauses

Inserting Follow the procedure below to insert a programmed pause.

To insert a programmed pause:

Step	Action
1	From the Create screen, use the Circular Key Pad to select a PCR segment time or temperature parameter where you want to insert a programmed pause.
2	Press More.
3	Press Insert. The Insert screen appears.

To insert a programmed pause: (continued)

Step	Action		
4	From the Insert screen, press Pause . The Programmed Pause screen appears.		
	Define Programmed Pause		
	Start first pause at cycle 1 of 25. Pause every 25 cycles for 00:30. Beep during the pause? No		
	Accept		
	F1 F2 F3 F4 F5		
5	In the Start first Pause at Cycle field, type in the cycle number where you want the method to first pause.		
6	In the Pause Every field, type in the pause frequency in cycles.		
	The pause frequency specifies the number of cycles that will run between each pause.		
7	In the Cycles For field, type in the length of the pause in minutes:seconds (00:01–98:59) format.		
8	In the Beep During The Pause? field, press Yes or No.		
9	Press Accept to accept the pause information on the screen.		
	The word Pause now displays to the right of the incubation step where you programmed the pause.		
	Note You can only insert one pause in each cycle.		

Editing If you have inserted a programmed pause in your method, you can edit the Programmed Pauses parameters for the pause at any time.

To edit programmed pauses:

Step	Action
1	From the Create screen, use the Circular Key Pad to highlight the word Pause . The Edit soft key appears.
2	Press Edit to access the programmed pause screen.
	From this screen, you can
	◆ Change any of the pause parameters, or
	♦ Use the Circular Key Pad to select the pause time parameter on the screen, and edit it by entering a different time.
	After you have entered all modifications to the customized method you are creating, you should store the method before running it (see "Naming and Storing Methods" below).

Methods

Naming and Storing Naming and storing completes the creation of the method.

To name and store methods:

Step	Action				
1	From the Create screen, press Store.				
2	You can take the following action:				
	If you want to Then				
	store the method under the name displayed in the user field and name the method the default name displayed in the method field	press the Accept key.			
	The default method name is expxxx where xxx= a number from 0 to 999.				
	rename the method	go to step 3.			
3	From the Store screen, press Method . The Method Name screen appears.				
	Note The blank space after the letter Z is used to insert blank spaces.				
	abcdefghi jklmnopqrj Method Name exp001 stuvwxyz				
	.,-+/():=				
	Use ENTER key to select a character.				
	[Accept] [Backsp] [Cancel] F1 F2 F3 F4 F5				
	F1 F2 F5 F4 F5				

To name and store methods: (continued)

Step	Action			
4	In the Method Name field, follow these steps to enter a 1–16 character alphanumeric name.			
	Step	Action		
	a.	Use the Circular Key Pad to select a character in the list shown in the upper right portion of the screen.		
	b.	After selecting a character, press Enter to place the character in the Method Name field.		
	C.	If you want to	Then	
		enter a number	press the appropriate number key.	
		go back one space	press the Backsp key.	
		clear the method name	press the CE key.	
5	Press Accept after you have entered a method name.			
	The Store screen appears again. The method name you entered should now display in the Method Name field.			
6	From the Store screen, press Accept.			
	This stores the method under the name you entered.			

Printing a Method

Introduction If you have configured your instrument for a printer, you can print a record of the parameters in a method. For more information on configuring a printer, see "Setting Custom Parameters" on page 3-8.

Printing a Method To print a method:

Step	Action
1	Access the Create or Edit screen.
	From the Create or Edit screen you can print a copy of the parameters for the method displayed on the screen.
2	Press Print .

Editing or Deleting Methods

Introduction After you create a method, you can edit its parameters, and store the method by the same name, or change its name. At some time, you may also want to delete a method if you are no longer using it. You can access all editing functions from the Main menu. The delete screen is accessed through the utility menu.

Editing a Method The following procedure describes how to edit a method.

To edit a method:

Action			
From the Main menu, press Edit .			
Note If an (SRAM) PC Card is detected in the Card slot, you will get a choice of editing a method on the PC Card or the instrument.			
The top line of the display continuously cycles between the following three lines:			
Methods on Inst User Size Stored [or Last Used] [or on PC card]			
Used Mem: xxx meth	ods xxx segments		
Free Mem: xxx metho	ods xxx segments		
The following table describe	es these fields.		
Field Description			
units for the Size field	Based on a calculation of the complexity and length of a method relative to a maximum size of 1102 size segments for the storage capacity of the instrument.		
Used Mem field Displays the number of segments used by all stormethods.		sed by all stored	
Free Mem field	Displays the number of segments a created methods.	vailable to store	
Select one of the methods follows:	displayed on the screen, or select and	other method as	
If you want to		Then press	
view the parameters of a method before making a selection		the View key.	
(Refer to "Viewing Method Parameters" on page 4-4.)			
search for a method by user name		the User key.	
(Refer to "Searching for Methods" on page 4-4.)			
sort methods by different of	criteria	the Sort key.	
(Refer to "Sorting Methods	s" on page 4-5.)		
	From the Main menu, press Note If an (SRAM) PC Cediting a method on the PC The top line of the display of Methods on Inst Use [or on PC card] Used Mem: xxx methods following table described Field units for the Size field Used Mem field Free Mem field Select one of the methods follows: If you want to view the parameters of a recommendation (Refer to "Viewing Methods search for a method by use (Refer to "Searching for Methods by different of sort methods by different of the sort method of the sort method of the sort method of the sort methods by different of the sort method of the sort	From the Main menu, press Edit. Note If an (SRAM) PC Card is detected in the Card slot, you we editing a method on the PC Card or the instrument. The top line of the display continuously cycles between the follow Methods on Inst User Size Stored [or Last Used] [or on PC card] Used Mem: xxx methods xxx segments Free Mem: xxx methods xxx segments The following table describes these fields. Field Description units for the Size field Based on a calculation of the complof a method relative to a maximum segments for the storage capacity of a methods. Free Mem field Displays the number of segments unethods. Free Mem field Displays the number of segments a created methods. Select one of the methods displayed on the screen, or select and follows: If you want to view the parameters of a method before making a selection (Refer to "Viewing Method Parameters" on page 4-4.) search for a method by user name	

To edit a method: (continued)

Step	Action		
3	Press Edit after selecting a method. The Edit screen appears.		
	2 Pre-PCR 3 Tmp 25 Cycles 2 Holds 94.0 94.0 55.0 10:00 0:30 55.0 72.0 2:00 55.0 0:30 5:00 Hold Cycle Pause Cancel		
4	F1 F2 F3 F4 F5		
4	Choose a temperature or time parameter within a PCR segment.		
5	Edit temperature control or time parameters.		
	Note Editing parameters on the Create screen involves the same tasks and uses the same key combinations as you use when creating a method. The same functions for modifying methods are also available.		
6	From the Edit screen, press Store to store the method.		

Deleting a Method The following procedure describes how to delete a method.

To delete a method:

Step	Action		
1	From the Utilities screen, screen appears.	press More and then press Delete. The	Delete Method
		ete Method st User Size Stored lisa 15 01/01/00	
	Press Yes to del		
	F1 F2	F3 F4 F5	
	Note If a PC Card is de method on the PC Card of	etected in the Card slot, you will get a char or the instrument.	oice of deleting a
	The top line of the display continuously cycles between the following three lines: Methods on Inst User Size Stored [or Last Used] [or on PC card]		
	Used Mem: xxx methods xxx segments		
	Free Mem: xxx methods xxx segments The following table describes these fields.		
Field Description			
	Units for Based on a calculation of the complexity and leng the Size field a method relative to a maximum size of 1102 size segments for the storage capacity of the instrume		
Used Mem Field Displays the number of segments used by all s methods.			d by all stored
	Free Mem Field	Displays the number of segments ava created methods.	ilable to store
2	Select one of the methods displayed on the screen, or select another method as follows:		
	If you want to		Then press
view method parameters (Refer to "Viewing Method Information" on page 4-8.) sort methods by different criteria (Refer to "Sorting Methods" on			the View key.
			the Sort key.
	page 4-5.)	t criteria (Neier to Sorting Methods on	the 3011 key.
3	Press Delete . The Delete	Confirmation screen appears.	
	If the method is protected is correct.	, enter a four-digit PIN and press Accept	when the number

To delete a method: (continued)

Step	Action	
4	Press Yes to confirm the deletion. This deletes the method and returns you to the Delete screen.	
	Note Even after you delete the last method stored under a User name, the name is removed from the instrument. To delete the name, see "Deleting a User Name" on page 5-5.	

Overview

About This Chapter

This chapter provides information about setting hold times for the GeneAmp® PCR System 9700 compared to the DNA Thermal Cycler or DNA Thermal Cycler 480. This information is based upon differences in how the instruments heat and cool samples.

Note Protocols using sample volumes between 5 μ L and 50 μ L, developed on either the GeneAmp® PCR System 2400 or 9600, may be transported to the GeneAmp PCR System 9700 without change when using the 9600 emulation mode.

In This Chapter The following topics are covered in this chapter:

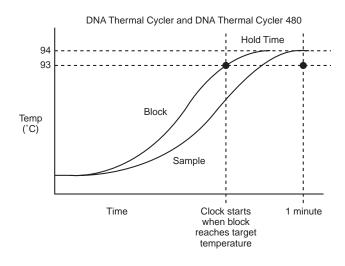
Topic	See page
About Setting Hold Times	6-2
Guidelines for Converting Hold Times	6-3

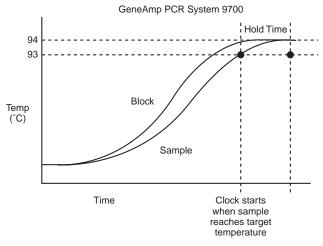
About Setting Hold Times

The hold times specified for the GeneAmp PCR System 9700 are shorter than those used for the Step-Cycle or ThermoCycle files on the DNA Thermal Cycler and the DNA Thermal Cycler 480. This is because the DNA Thermal Cycler and the DNA Thermal Cycler 480 starts counting the hold time when the block reaches a temperature one degree before the target temperature, while the GeneAmp PCR System 9700 starts counting the hold time when the samples reach a temperature one degree before the target temperature.

Hold Time Differences

As shown in the figures below, since the block reaches the target temperature before the sample, the programmed hold time on the DNA Thermal Cycler and the DNA Thermal Cycler 480 must include enough time for the samples to reach the target temperature. A hold time of one minute or greater is required on the DNA Thermal Cycler and the DNA Thermal Cycler 480 for samples to reach the target temperature. On the GeneAmp PCR System 9700, hold times of less than one minute are generally used.





Guidelines for Converting Hold Times

Introduction This section describes guidelines on how to convert hold times for the DNA Thermal Cycler or the DNA Thermal Cycler 480 to hold times for the GeneAmp PCR System 9700 when using the 9600 mode.

New Hold Times On or down ramp.

What to Base the The following table lists what to base the new hold time on if you are using an up ramp

If you are using	Then base the new hold time	For more information see
an up ramp	on the change in the temperature required to reach the next target temperature.	Table 6-1 on page 6-3.
a down ramp	on the starting temperature of the ramp and the change in temperature required to reach the next temperature.	Table 6-2 on page 6-5.

Setting Up Ramps The following table lists the process of setting the up ramp temperature.

Step	Action	
1	Determine the change in temperature required to reach the next target temperature and round this value off to the closest value found in Table 6-1.	
2	Subtract the number of seconds indicated from the hold time used for the DNA Thermal Cycler or the DNA Thermal Cycler 480.	
	The result is the hold time to use for the GeneAmp PCR System 9700.	
	Note The typical hold time is 10 to 15 seconds for denaturation.	

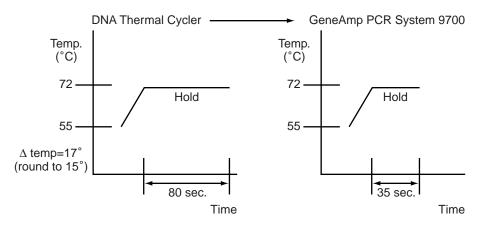
Table of Conversions The following table lists the times for converting up ramp hold times greater than one minute from the DNA Thermal Cycler and the DNA Thermal Cycler 480 to the GeneAmp PCR System 9700.

Table 6-1 Converting up ramp hold times

∆ Temp (°C)	Seconds to subtract from DNA Thermal Cycler or DNA Thermal Cycler 480 hold times (>1 min.)
10°	38 sec.
15°	45 sec.
20°	49 sec.
30°	54 sec.
40°	55 sec,
50°	57 sec.
60°	57 sec.

Up Ramp Example In this example, the temperature was increased by 17 °C. This value was rounded to 15 °C. According to Table 6-1, subtract 45 seconds from the hold time on the DNA Thermal Cycler or the DNA Thermal Cycler 480, resulting in a new hold time of 35 seconds (see the figure below).

> Note If methods developed on the GeneAmp PCR System 9700 will be used on the DNA Thermal Cycler or the DNA Thermal Cycler 480, you can also use Table 6-1 to convert the hold times. Add the indicated times instead of subtracting them.



Setting Down Ramps The following table lists the process of setting the down ramp temperature.

Step	Action	
1	Determine the change in temperature required to reach the next target temperature and round this value off to the closest value found in Table 6-2.	
2	Based on the starting temperature of the ramp, determine the number of seconds to subtract from the DNA Thermal Cycler or DNA Thermal Cycler 480 hold time to arrive at the new hold time.	
	Note The typical hold time is 10 to 15 seconds for annealing.	

Table of Conversions

The following table lists the times for converting down ramp hold times greater than one minute from the DNA Thermal Cycler or the DNA Thermal Cycler 480 to the GeneAmp PCR System 9700.

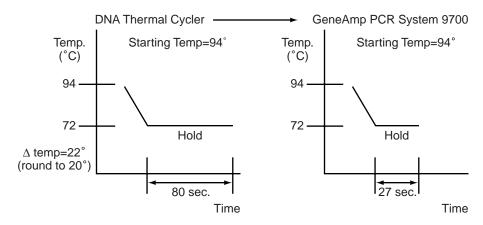
Table 6-2 Converting down ramp hold times

	Starting temperature of ramp (°C)		
A Temp (°C)	35-55°	56-75°	76-96°
10°	39 sec.	39 sec.	40 sec.
15°	45 sec.	47 sec.	48 sec.
20°	49 sec.	52 sec.	53 sec.
30°	_	56 sec.	59 sec.
40°	_	57 sec.	62 sec.
50°	_	_	62 sec.
60°	_	_	60 sec.

Down Ramp **Example**

In the example below, the temperature was decreased by 22 °C. This value was rounded to 20 °C. According to Table 6-2, with a starting temperature of 94 °C, we should subtract 53 seconds from the hold time on the DNA Thermal Cycler or the DNA Thermal Cycler 480, resulting in a new hold time of 27 seconds.

Note If the methods developed on the GeneAmp PCR System 9700 will be used on the DNA Thermal Cycler or the DNA Thermal Cycler 480, use Table 6-2 to convert the hold times. Add the indicated times instead of subtracting them.



Routine Maintenance

Overview

About This Chapter

This chapter describes how to perform routine maintenance on the GeneAmp® PCR System 9700.

A CAUTION Do not remove the instrument cover. There are no components inside the GeneAmp PCR System 9700 that you can safely service yourself. If you suspect a problem, contact an Applied Biosystems Technical Support Representative.

In This Chapter The following topics are covered in this chapter:

Торіс	See page
Changing the External Fuses	7-2
Changing Fuses for Single-Line Operation	7-3
Changing Fuses for Double-Line Operation	7-4

Changing the External Fuses

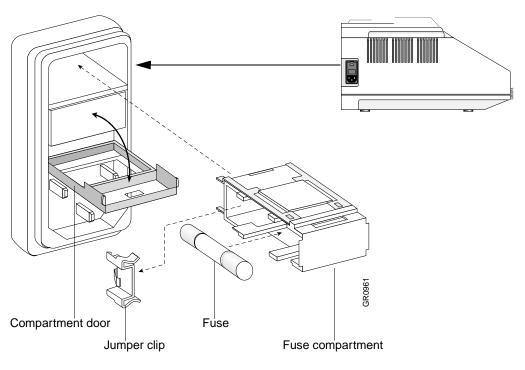
Introduction

A CAUTION FIRE HAZARD. For continued protection against the risk of fire, replace fuses only with listed and certified fuses of the same type and rating as those currently in the instrument.

All instruments have factory installed fuses. However, if you should ever need to change a fuse, or if you need to convert from the factory configured single-line service to double-line service, follow the instructions that follow in this chapter.

Power Entry Module Diagram

The Power Entry Module is located at the rear of the left side of the instrument.



Single-Line Operation

Changing Fuses for There is one Type T 250 V fuse accessible from the side of the instrument. You need a flat-tip screwdriver to open the door of the Power Entry Module and change the fuse.

To change the fuse for single-line operation:

Step	Action		
1	Turn off the instrument and disconnect the power cord from the side of the instrument.		
	A DANGER ELECTRICAL SHOCK HAZARD. Severe electrical shock, which could cause physical injury or death, can result from working on an instrument when the high voltage power supply is operating. To avoid electrical shock, disconnect the power supply to the instrument, unplug the power cord, and wait at least 1 minute before working on the instrument.		
	The fuse is located in the Power Entry Module.		
2	Insert a small flat-tip screwdriver into the slot in the upper portion of the power entry module, and open the door.		
3	Pull the fuse compartment out. There is one fuse in the fuse compartment.		
4	Pull out the fuse from the back of the fuse compartment and replace it with either:		
	♦ an 8 Amp Type T (P/N 0999-1683) or,		
	♦ a Slow-Blow 250 V fuse (P/N 0998-1643).		
5	Place the fuse compartment back into the Power Entry Module and close the door.		
	Press the door until it locks in place.		
6	Connect the instrument power cord.		

Operation

Changing Fuses for Instruments configured for double-line operation have two 8 amp Type T 250 V **Double-Line** 5x20 mm fuses (P/N 0999-1683).

To change the fuses for double-line instruments:

Step	Action
1	Turn off the instrument and disconnect the power cord from the side of the instrument.
	⚠ DANGER ELECTRICAL SHOCK HAZARD. Severe electrical shock, which could cause physical injury or death, can result from working on an instrument when the high voltage power supply is operating. To avoid electrical shock, disconnect the power supply to the instrument, unplug the power cord, and wait at least 1 minute before working on the instrument.
	The fuses are located in the Power Entry Module.
2	Insert a small flat-tip screwdriver into the slot in the upper portion of the power entry module, and open the door.
3	Pull the fuse compartment out.
	There are two fuses in the fuse compartment.
4	Pull out the fuse from the back of the fuse compartment and replace the blown fuse with one 8 amp Type T 250 V fuse.
5	Place the fuse compartment back into the Power Entry Module and close the door.
	Press the door until it locks in place.
6	Connect the instrument power cord.

Troubleshooting

Overview

About This Chapter

This chapter describes instrument problems you may have, the probable causes of these problems, and any display screen messages you may encounter when using the GeneAmp® PCR System 9700.

In This Chapter The following topics are covered in this chapter:

Topic	See page
If There Is a Power Failure	8-2
Display Screen Error Messages	8-3
Troubleshooting Information	8-6

If There Is a Power Failure

Introduction An automated restart function allows for power failures and safe continuation of a PCR run after resumption of power.

Failure

During a Power The following table lists the actions the instrument takes if the power is interrupted and the instrument turns off while you are operating it.

> Note If the power is off for 15 seconds or longer and fails during execution of a cycle then the cycle currently running will restart. If the power fails while executing a hold, or approaching a hold, then that hold temperature will restart from the beginning.

Note If the power failure lasts longer than 18 hours, the Resume will not occur.

Do the following in a power failure:

Step	Action
1	Restart or continue the PCR experiment.
	The instrument determines what temperature was being approached, or was holding.
	Upon resumption of power, it will go to that temperature and countdown the time remaining in the hold as soon as the temperature is within the specified clock start limits.
2	Incubate the samples until you can continue the experiment.
3	Enter a record for any power outage in the history file.

Display Screen Error Messages

Error Messages Refer to the following table for a description of error messages, and recommended Table actions that you should take.

Table 8-1 Error Messages

Message	Description	Recommended Action
Battery RAM version number lost	RAM version number This error is generated when the battery RAM has been lost and re-initialized.	
Block Calibration initialized	Software or hardware failure.	Call Technical Support.
Block Calibration reset to	System error.	Call Technical Support.
default	Block data reset to defaults.	
Block isn't configured	The instrument has defaulted to a 96-well configuration.	Call Technical Support.
Block initialized	Block module has been re-initialized.	Call Technical Support.
Block version unknown, update firmware	The calibration data in the block is not recognized by the firmware.	Upgrade the firmware.
Bus Error	System error.	Call Technical Support.
Calibration battery RAM initialized	Calibration lost. Instrument may not perform to specification.	Call Technical Support.
Can only enter an infinity hold at end	A method can only have a HOLD segment with an infinity hold as the last segment in a method.	Assign finite time segments to holds within a method.
	This message occurs when you try to enter an infinity hold segment in the middle of a method.	
Can't allocate timer	System error.	Call Technical Support.
Delete your methods first	User tried to delete a user name that has methods stored under it.	Delete or transfer the associated methods before deleting a user name.
Enter a name or CANCEL	You did not enter at least one character on the User Name screen before pressing the Accept key.	Enter the user name to which the desired method is assigned.
Enter oligo sequence	Incomplete TmCalc data.	Enter a value in the P1P2 fields of the TmCalc.
Enter user and method names or CANCEL	You did not enter a user name and a method name before storing a method.	Specify the method name and choose a user to store a method.
FATAL – Block shut off by hardware	Block thermal runaway.	Call Technical Support.
FATAL – Block thermal runway	Fatal error.	Turn off system.
		Call Technical Support.
FATAL – Cover shut off by hardware	Heated cover thermal runaway.	Call Technical Support.
FATAL – Heat sink is too hot	Ambient conditions may be too warm.	Call Technical Support.
FATAL – Heat sink sensor failure	System error.	Call Technical Support.
FATAL – Heated cover thermal	Fatal error.	Turn off system.
runaway		Call Technical Support.

 Table 8-1
 Error Messages (continued)

Message	Description	Recommended Action
FATAL – Sample block sensor	Fatal error.	Turn off system.
failure		Call Technical Support.
Fatal – Stack Overflow	A warning or error message that displays which task stack overflowed.	Call Technical Support.
	The warning message is issued when the stack has reached within 10% of overflowing.	
Field is full	You tried to enter more data in a field than the field size allows.	Reenter data within the specifications of the field.
Heated cover sensor failure	The heated cover sensor failed.	Call Technical Support.
Infinity hold not allowed in cycle	A method can have an infinity hold segment as the last segment in the method.	Use finite values for cycle segments within the method.
	This message occurs when you tried to enter an infinity time in a CYCLE segment.	metriod.
Invalid password/pin#	You entered an incorrect PIN#.	Enter the correct PIN#.
LCD screen timed-out	System error.	Call a Technical Support.
	Display screen and firmware have a faulty connection.	
List of user names is full	The maximum number of users has been entered into the system.	Delete unused user names.
Maximum of 6 segments allowed	You tried to insert more than six temperature control parameters into a hold or cycle.	Do not assign more than six hold or cycle parameters to a method.
Method battery RAM initialized	Stored methods have been reset due to hardware or software failure. Not all methods may be lost.	Check method directory. Call Technical Support.
Method requires at least one segment	You deleted all temperature control parameters in a method.	Review and correct the method to include the temperature parameter(s).
	A method must have at least one time and temperature parameter.	temperature parameter(s).
No seconds in time field	You did not include seconds in the time field.	Include seconds when entering the time.
Not enough method memory	This error occurs:	♦ Determine how much
left	When you attempt to exceed the limit of 137 methods.	storage memory is available on the instrument or PC card.
	♦ When you attempt to store or create a new method which is larger than the available storage space.	Delete or store rarely used methods elsewhere.
Not implemented yet	The feature is not implemented in the current firmware version.	Upgrade firmware when the new version is available from Applied Biosystems.
PC card and Flash do not verify	The PC (upgrade) card and instrument memory do not match.	Call Technical Support.
	Firmware upgrade unsuccessful.	
PC card does not contain valid data	The PC card being used to upgrade the instrument does not contain a valid program.	Call Technical Support.

 Table 8-1
 Error Messages (continued)

Message	Description	Recommended Action
Preferences battery RAM initialized	User configuration has been reset due to software error.	Call Technical Support.
Printer not responding	rinter not responding The printer has been disconnected or is off line.	
Remove infinity hold first	A method can have an infinity hold segment as the last segment in the method.	Add segments prior to the post-PCR infinity hold.
	This message occurs when a user tries to add a segment after one which contains an infinity hold.	
Setpoint could not be reached	◆ The instrument could not reach a temperature parameter set by the user.	Call Technical Support.
	The unit has a Peltier or power amplifier failure.	
	 Ambient conditions may be out of recommended range. 	
SYSTEM ERROR invalid pointer	System error.	Call Technical Support.
Tm temperature out of range	Tm out of range.	Check input value and retry.
		Call Technical Support.
User name already defined	You entered a user name that already exists.	Do not duplicate user names.
WARNING: Block version	Some data in the block is unrecognized.	Upgrade firmware.
unknown	Instrument operation will not be effected.	
Watchdog timeout	Software failure.	Call Technical Support.
Write to block failed	Information written to the Interchangeable Sample Block Module has failed.	Call Technical Support.
Write to default block failed	System error.	Call Technical Support.
	Write to memory in block failed.	
Write to default Xicor failed	System error.	Call Technical Support.
	Write to memory in the block failed.	
Write to Xicor failed	Information written to the Interchangeable Sample Block Module has failed.	Call Technical Support.
Valid range is	You entered a number out of range.	Reenter a value within the
	The message include the valid range limits.	parameters of the field.

Troubleshooting Information

Troubleshooting Refer to the following table for a description of potential problems, possible causes, Table and recommended actions that you should take.

 Table 8-2
 Troubleshooting Information

Problem	Possible Causes	Check and/or Remedy
Control panel not responding	Keypad failure.	Run keypad diagnostic.
		Call Technical Support.
Cooling rate too slow	Ambient temperature is too warm.Peltier failure.	♦ Move instrument to well-ventilated location(15-30°C).
		♦ Run rate test diagnostic.
		Call Technical Support.
Cycling time too long	Peltier failure.	Run cycle test diagnostic
Displayed temperature does not match specified temperature	Instrument may require calibration.	Run the Temperature Verification test.
Heated cover not responsive	Heated cover failure.	Call Technical Support.
Heating rate too slow	→ Peltier failure.	Run Rate Test diagnostic.
		Call Applied Biosystems Technical Support.
Instrument can't reach high or low	♦ Ambient temperature is too warm.	◆ Run Rate Test diagnostic.
temperature range	◆ Peltier failure.	◆ Run Cycle Test diagnostic.
		Call Technical Support.
Instrument making too much noise	Fan failure.	Check for sidevent obstructions.
No beep	Run time beeper disabled.Beeper failure.	Check Run-Time Beep configuration.
	V Beeper landre.	Call Technical Support.
No screen display	♦ Fuse blown.	♦ Is power switch ON?
No response when you turn the	♦ Not connected to power source.	♦ Is power cord connected?
instrument on	 Interchangeable module not installed correctly. 	♦ Check fuses.
Printer fails	♦ Incorrect printer configuration.	◆ Check printer settings: baud
	♦ Incorrect printer cable.	rate = 9600, no parity, one stop bit, eight data bits.
		 Purchase Applied Biosystems printer cable.
Instrument cooling fan does not make	♦ Fuse blown.	♦ Is power switch ON?
whirring sound	♦ Not connected to power source.	♦ Is power cord connected?
	 Interchangeable module not installed correctly. 	♦ Check fuses.

Instrument Specifications



Overview

About This This appendix describes the dimensions, power, and electrical specifications for the Appendix GeneAmp® PCR System 9700 system, including the control panel, sample temperature information, and printer specifications.

In This Appendix The following topics are covered in this appendix:

Topic	See page
System Specifications	A-2
Control Panel Specifications	A-3
Sample Temperature Information	A-3
Printer Specifications	A-3

System Specifications

The following tables list the footprint and the weight of the instrument with the 96-Well or 60-Well sample block module.

Footprint (With Sample Block Module Installed)

Height	26 cm (10 in)
Width	28 cm (11 in)
Depth	41 cm (16 in)

Note You must provide sufficient space around the instrument for unrestricted air circulation.

Weight

Base Module	8.6 kg (19 lbs)
96-Well Gold/Aluminum Sample Block Modules	3.2 kg (7 lbs)

Note See the sample block module user's manual for physical information on a module.

Configurations

Power There is one version of the instrument. The power requirements of the instrument under various power configurations are:

VAC ~100/120	8 AMP T (5x20 mm) or	50/60 Hz
	8 AMP Slow Blow (3 AB)	Use 250 V fuses
VAC ~220/230/240	8 AMP T (5x20 mm)	Max Power 725 VA

Requirements

Electrical IMPORTANT You must be able to disconnect the main power supply to the instrument immediately if necessary.

> The following table specifies the electrical operating range for the instrument in various parts of the world. Select appropriate fuse configuration based on the voltage used.

A WARNING In Japan, the unit must have a dedicated 220-volt outlet! The unit will not operate properly with a 100-volt outlet.

Location	Voltage (VAC) ^a	Frequency	Amperage (A) Nominal
Japan	220 ±10%	50/60 Hz ±1%	3.16
USA/Canada	120 ±10%	50/60 Hz ±1%	4.20
Europe (pre-1992)	220 ±10%	50/60 Hz ±1%	3.16
EC	230 ±10%	50/60 Hz ±1%	3.14
UK (pre-1992)	240 +6%/–10%	50/60 Hz ±1%	3.12
Australia	240 +6%/–10%	50/60 Hz ±1%	3.12

a. Acceptable AC line voltage tolerances: 100, 120, 220, 230 \pm 10%; 240 VAC \pm 6%/ \pm 10%, 50/60 Hz \pm 1%.

Note The Volt-Amp number for this instrument is 725Volt Amps.

Control Panel Specifications

Display Screen The display screen is a 7 x 40 character display with a graphics mode of 60 x 240 pixel resolution.

Keys The instrument control panel consists of a display screen and 22 keys. The keys are:

- Function keys
- Arrow keys
- Stop key
- Enter key
- 10 number keypad

Sample Temperature Information

Temperature The following table lists sample temperature information.

Note Sample temperatures are displayed in degrees Celsius to the nearest 0.1 °C.

Sample Temperature Range	4.0 to 99.9 °C.
Temperature Calibration	Traceable to National Institute of Standards and Technology (NIST).

Printer Specifications

Board Specifications following parameters.

Serial Interface The instrument can use any printer with a serial (RS-232C) interface board with the

Baud Rate	9600
Parity	NONE
Data Bits	8
Stop Bits	1

Cable Part Number Connect the printer to the Instrument port with printer cable part number N805-1326.

Supplied Methods

Overview

About this Appendix This appendix provides information about the methods that are supplied with the GeneAmp® PCR System 9700.

In This Appendix The following topics are covered in this appendix:

Topic	See page
About the Methods	B-2
AmpliCycle Sequencing	B-2
AmpliTaq Gold	B-2
BigDye	B-3
General PCR	B-3
LMS2	B-4
Time Release PCR	B-4
Touchdown PCR	B-5
XL PCR	B-5

About the Methods

Introduction

The instrument supplies you with eight pre-coded methods stored under the user name <<pe>>>.

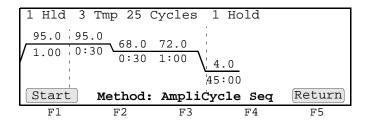
The eight pre-coded methods are:

- AmpliCycle® Sequencing
- AmpliTaq Gold®
- BigDye™
- General PCR
- LSM2
- Time Release PCR
- Touchdown PCR
- XL PCR

See the following sections for detailed information about the methods.

AmpliCycle Sequencing

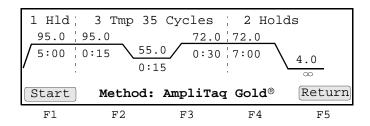
Cycle sequencing has revolutionized Sanger sequencing of PCR products and other DNA templates. The AmpliCycle Sequencing protocol is for 3-temperature cycling and for achieving clean sequence ladders from femtomole amounts of template.



This cycle sequencing process and the benefits of AmpliTag® DNA Polymerase, CS, are described in the product insert for the AmpliCycle® Sequencing Kit (P/N N808-0175).

AmpliTaq Gold

The AmpliTag Gold protocol specifies a 5-minute pre-PCR heat step, required for the activation of AmpliTaq Gold® DNA Polymerase. This additional step provides seamless "hot start" PCR and replaces labor intensive methods such as manual hot start or wax bead-mediated hot start techniques.

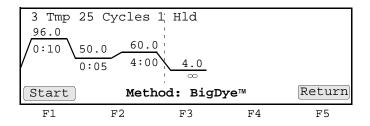


Utilizing hot start techniques helps to minimize the formation of primer-dimers or non-specific products, thus increasing specificity and sensitivity of PCR.

You can find further information on AmpliTaq Gold DNA Polymerase in the product insert (P/N N808-0241) or at the Applied Biosystems website.

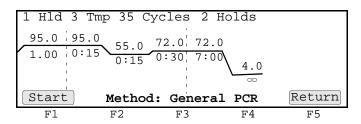
BigDve

The BigDye™ method consists of cycle sequencing parameters for dideoxy (Sanger) terminator sequencing using ABI PRISM® BigDye™ Terminator Cycle Sequencing Ready Reaction Kits (available from Applied Biosystems). It consists of 25-cycle. three-temperature cycle sequencing followed by an infinite hold at 4 °C.

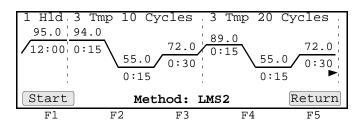


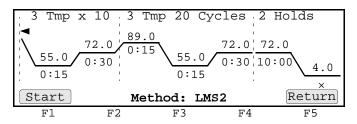
The pre-coded conditions in the BigDye™ method are optimized for AmpliTag® DNA Polymerase, FS and the cycle sequencing reagents supplied with the BigDye terminator kits. This process is further described in the ABI PRISM BigDye Terminator Cycle Sequencing Ready Reaction Kits Protocol (P/N 4303237).

General PCR The General PCR method is a basic one and can be easily modified with both preand post-PCR holds.



LMS2 The GeneAmp PCR System 9700 software includes a pre-coded LMS2 method for ABI PRISM® Linkage Mapping Set Version 2 (LMS2) thermal cycling. Linkage Mapping Set 2 employs over 400 fluorescent-labeled PCR primer pairs for analysis of select microsatalite loci from the Généthon human linkage map. 1,2,3 The following figures illustrate the thermal cycling profile for the LMS2 method.





This supplied method consists of an initial hold at 95.0 °C, two sets of three temperature cycles, followed by two additional holds at 72.0 °C and 4.0 °C.

See the ABI PRISM Linkage Mapping Set Version 2 User's Manual (P/N 904999) for comprehensive information on PCR amplification conditions, electrophoresis conditions, detection, and data analysis.

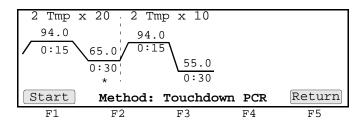
Time Release PCR The Time Release PCR method is designed for use with AmpliTaq Gold DNA Polymerase. The enzyme is activated more slowly than with the AmpliTaq Gold method. Here the pre-PCR hold is only 1 minute, and the number of cycles is increased to 40.

^{1.} Weissenbach, J. et al. 1992. A second-generation linkage map of the human genome. Nature 359:794-801.

^{2.} Gyapay, G., et al. 1994. Généthon Human Genetic Linkage Map. Nature Genet. 7:246-339.

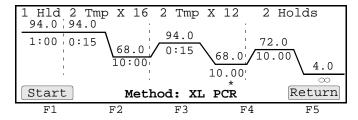
^{3.} Dib, C., et al. 1996. Nature 380: 152-154.

Touchdown PCR When the optimal annealing temperature is unknown, one strategy, touchdown PCR, incrementally decreases the annealing temperature in early cycles in order to maximize the yield of specific products.



This supplied method has an initial annealing temperature (65 °C) that incrementally decreases by an additional 0.5 °C in each of the first 20 cycles, followed by 10 cycles at 55 °C.

XL PCR XL PCR is the protocol specified for amplification of 5 kb-40 kb PCR products, using rTth DNA Polymerase, XL, and unique reaction conditions.



This protocol uses two-temperature cycling (94 °C for 15 seconds; 68 °C for 10 minutes) and invokes a 15 second AutoX (automatic segment extension) for the anneal/extend step in the last 12 cycles.

By providing longer templates, XL PCR complements technologies for rapid, long-range PCR. More complete genes can be amplified in one reaction from known expressed sequences, thus more introns can be crossed. You can use XL PCR for the amplification of the control target, a 20.8 kb product from Lambda DNA, supplied in the kit.

This process is further described in the product insert for the GeneAmp® XL PCR Kit (P/N N808-0192).

Contacting Technical Support



Technical Support

Technical Support

Contacting You can contact Applied Biosystems for technical support:

- By e-mail
- By telephone or fax
- Through the Applied Biosystems web site

You can order Applied Biosystems user documents, MSDSs, certificates of analysis, and other related documents 24 hours a day. In addition, you can download documents in PDF format from the Applied Biosystems web site. (Please see the section "To Obtain Technical Documents" following the telephone information below)

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Product/Product Area	E-mail address
Genetic Analysis (DNA Sequencing)	galab@appliedbiosystems.com
Sequence Detection Systems (Real-Time PCR) and PCR	pcrlab@appliedbiosystems.com
Protein Sequencing, Peptide, and DNA Synthesis	corelab@appliedbiosystems.com

Product/Product Area	E-mail address
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 MassGenotyping Solution 1™ (MGS1) Systems 	
◆ PNA Custom and Synthesis	
♦ Pioneer™ Peptide Synthesizers	
◆ Proteomics Solution 1 [™] (PS1) Systems	
♦ ICAT™ Reagent	
♦ FMAT™ 8100 HTS Systems	
 Mariner™ ESI-TOF Mass Spectrometry Workstations 	
♦ Voyager™ MALDI-TOF Biospectrometry Workstations	
◆ CytoFluor® 4000 Fluorescence Plate Reader	
LC/MS (Applied Biosystems/MDS Sciex)	support@sciex.com
Chemiluminescence (Tropix)	tropix@appliedbiosystems.com

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Note To schedule a service call for other support needs, or in case of an emergency, dial (North America) 1.800.831.6844, then press 1.

Product/Product Area	Telephone	Fax
ABI PRISM® 3700 DNA Analyzer	1.800.831.6844, then press 8 ^a	1.650.638.5981
DNA Synthesis	1.800.831.6844 , press 2 , then press 1 ^a	1.650.638.5981
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Fluorescent Fragment Analysis (including GeneScan® applications)	1.800.831.6844 , press 2 , then press 3 ^a	1.650.638.5981
Integrated Thermal Cyclers (ABI PRISM® 877 and Catalyst 800 instruments)	1.800.831.6844 , press 2 , then press 4 ^a	1.650.638.5981
ABI PRISM® 3100 Genetic Analyzer	1.800.831.6844 , press 2 , then press 6 ^a	1.650.638.5981
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Protein Sequencing (Procise® Protein Sequencing Systems)	1.800.831.6844 , press 3 , then press 2 ^a	1.650.638.5981
Sequence Detection Systems (Real-Time PCR) and PCR	1.800.762.4001, then press:	1.240.453.4613
	1 for PCR ^a	
	2 for TaqMan® applications and Sequence Detection Systems including ABI Prism 7700, 7900, and 5700a	
	6 for the 6700 Automated Sample Prep System ^a	
	or 1.800.831.6844, then	
	press 5 ^a	
Mariner™ ESI-TOF Mass Spectrometry Workstations	1.800.899.5858 , press 1 , then press 3 ^b	1.508.383.7855
♦ Voyager™ MALDI-TOF Biospectrometry Workstations		
 MassGenotyping Solution 1™ (MGS1) Systems 		
◆ Proteomics Solution 1 [™] (PS1) Systems		
◆ ICAT™ Reagent		

Product/Product Area	Telephone	Fax
Biochromatography (BioCAD®, SPRINT™, VISION™, and INTEGRAL® Workstations and POROS® Perfusion Chromatography Products)	1.800.899.5858 , press 1 , then press 4 ^b	1.508.383.7855
Expedite™ 8900 Nucleic Acid Synthesis Systems	1.800.899.5858, press 1, then press 5 ^b	1.508.383.7855
Pioneer™ Peptide Synthesizers	1.800.899.5858 , press 1 , then press 5 ^b	1.508.383.7855
PNA Custom and Synthesis	1.800.899.5858 , press 1 , then press 5 ^b	1.508.383.7855
 ► FMAT™ 8100 HTS Systems ► CytoFluor® 4000 Fluorescence Plate Reader 	1.800.899.5858 , press 1 , then press 6 ^b	1.508.383.7855
Chemiluminescence (Tropix)	1.800.542.2369 (U.S. only), or 1.781.271.0045°	1.781.275.8581
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China (Beijing)	86 10 64106608 or 86 800 8100497	86 10 64106617
Hong Kong	852 2756 6928	852 2756 6968
India (New Delhi)	91 11 653 3743/3744	91 11 653 3138
Korea (Seoul)	82 2 593 6470/6471	82 2 593 6472
Malaysia (Petaling Jaya)	60 3 79588268	60 3 79549043
Singapore	65 896 2168	65 896 2147
Taiwan (Taipei Hsien)	886 2 2358 2838	886 2 2358 2839
Thailand (Bangkok)	66 2 719 6405	66 2 319 9788
Europe		
Austria (Wien)	43 (0)1 867 35 75 0	43 (0)1 867 35 75 11
Belgium	32 (0)2 532 4484	32 (0)2 582 1886
Denmark (Naerum)	45 45 58 60 00	45 45 58 60 01
Finland (Espoo)	358 (0)9 251 24 250	358 (0)9 251 24 243
France (Paris)	33 (0)1 69 59 85 85	33 (0)1 69 59 85 00
Germany (Weiterstadt)	49 (0)6150 101 0	49 (0)6150 101 101
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Region	Telephone	Fax
Norway (Oslo)	47 23 12 06 05	47 23 12 05 75
Portugal (Lisboa)	351.(0)22.605.33.14	351.(0)22.605.33.15
Spain (Tres Cantos)	34.(0)91.806.1210	34.(0)91.806.12.06
Sweden (Stockholm)	46 (0)8 619 4400	46 (0)8 619 4401
Switzerland (Rotkreuz)	41 (0)41 799 7777	41 (0)41 790 0676
The Netherlands (Nieuwerkerk a/d IJssel)	31 (0)180 392400	31 (0)180 392409 or 31 (0)180 392499
United Kingdom (Warrington, Cheshire)	44 (0)1925 825650	44 (0)1925 282502
European Mar	naged Territories (EMT)	
Africa, English speaking (Johannesburg, South Africa)	27 11 478 0411	27 11 478 0349
Africa, French speaking (Paris, France)	33 1 69 59 85 11	33 1 69 59 85 00
India (New Delhi)	91 11 653 3743	91 11 653 3138
	91 11 653 3744	
Poland, Lithuania, Latvia, and Estonia (Warszawa)	48 22 866 40 10	48 22 866 40 20
For all other EMT countries not listed (Central and southeast Europe, CIS, Middle East, and West Asia)	44 1925 282481	44 1925 282509
	Japan	
Japan (Hacchobori, Chuo-Ku, Tokyo)	81 3 5566 6230	81 3 5566 6507
La	tin America	
Caribbean countries, Mexico, and Central America	52 55 35 3610	52 55 66 2308
Brazil	0 800 704 9004 or 55 11 5070 9654	55 11 5070 9694/95
Argentina	800 666 0096	55 11 5070 9694/95
Chile	1230 020 9102	55 11 5070 9694/95
Uruguay	0004 055 654	55 11 5070 9694/95

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	♦ Click the pdf icon to view a PDF version of the document.					
	◆ Right-click the pdf icon, then select Save Target As to download a copy of the PDF file.					
	♦ Select the Fax check box, then click Deliver Selected Documents Now to have the document faxed to you.					
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The Warranty does not extend to any Instrument or part thereof that has been the subject of misuse, neglect or accident, that has been modified or repaired by any party other than Applied Biosystems or its designees, or that has been used in a manner not in accordance with the instructions contained in the Instrument User's Manual.

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